



多重抗藥性細菌之抗藥機轉

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課程大綱

1. 抗生素對細菌之作用機轉

2. 細菌抗藥性機轉之演進

3. 抗藥性革蘭氏陽性菌之抗藥機轉

4. 抗藥性革蘭氏陰性菌之抗藥機轉

5. 結論



1. 抗生素對細菌之作用機轉

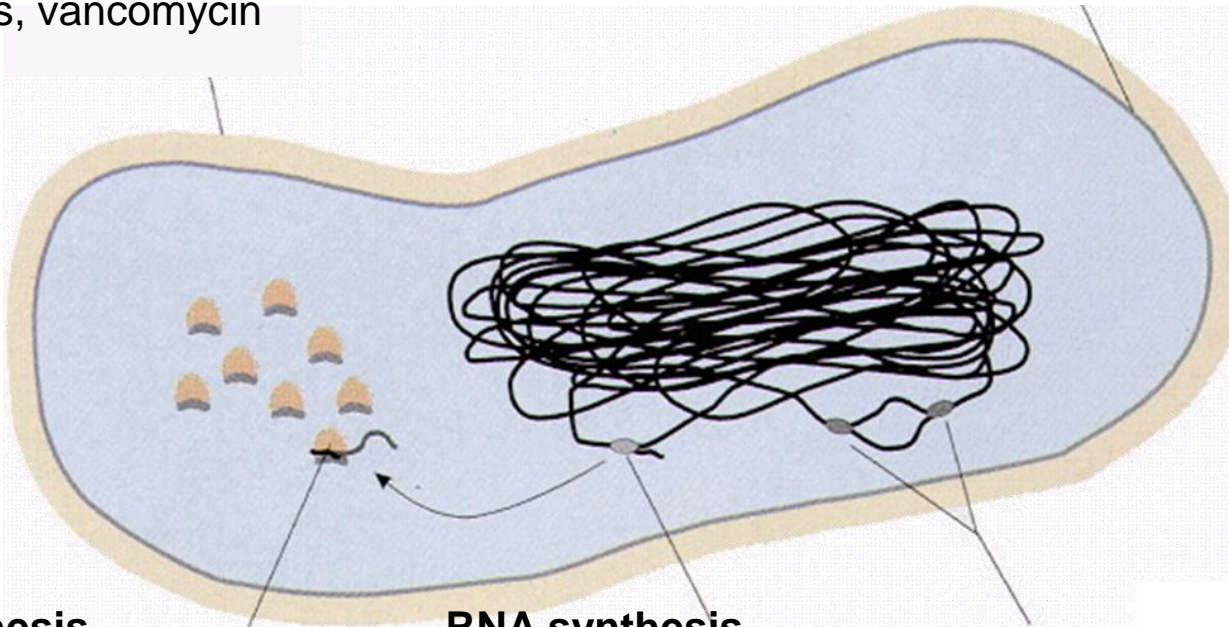
細菌的結構

Peptidoglycan cell wall

Some antibiotics weaken the wall, causing the cell to rupture-penicillins, cephalosporins, vancomycin

Plasma membrane

Some antibiotics make the membrane leaky, allowing essential small molecules to leak out of the cell-polymyxins



Protein synthesis

Some antibiotics inhibit ribosomes which synthesise the proteins required by the cell-aminoglycosides, tetracyclines, macrolides

RNA synthesis

Some antibiotics inhibit synthesis of the RNA molecules that carry the genetic code between the DNA and the ribosomes-rifampicin

DNA synthesis

Some antibiotics inhibit replication of the genetic code-quinolones, novobiocin

抗微生物製劑（抗生素）作用機轉

抑制細胞壁合成

- Penicillins, cephalosporins, glycopeptides, carbapenems

抑制蛋白質合成

- Aminoglycoside, macrolides, clindamycin, tetracyclines

抑制核酸合成

- Quinolones, rifampin

抑制細胞膜功能

- Polymyxins, daptomycin

抑制營養代謝及生長

- Sulfonamide

抑制細胞壁合成

- Cell wall polymer - peptidoglycan inhibitors
β-lactams : penicillins 青黴素
cephalosporins 頭孢子素
carbapenems 碳氫黴烯
- Block cross-linking of peptidoglycan
Glycopeptides : vancomycin 萬古黴素
teicoplanin; bacitracin
- Fosfomycin : inhibit MurA biogenesis

抑制蛋白質合成

- 抑制30S核糖體蛋白質合成(30s ribosome)
Aminoglycosides : gentamicin, tobramycin,
amikacin, streptomycin
- 抑制50S核糖體蛋白質合成(50s ribosome)
Macrolides : erythromycin, clarithromycin,
azithromycin, clindamycin,
chloramphenicol
- 抑制70S複合體形成(70s initiation complex)
Linezolid

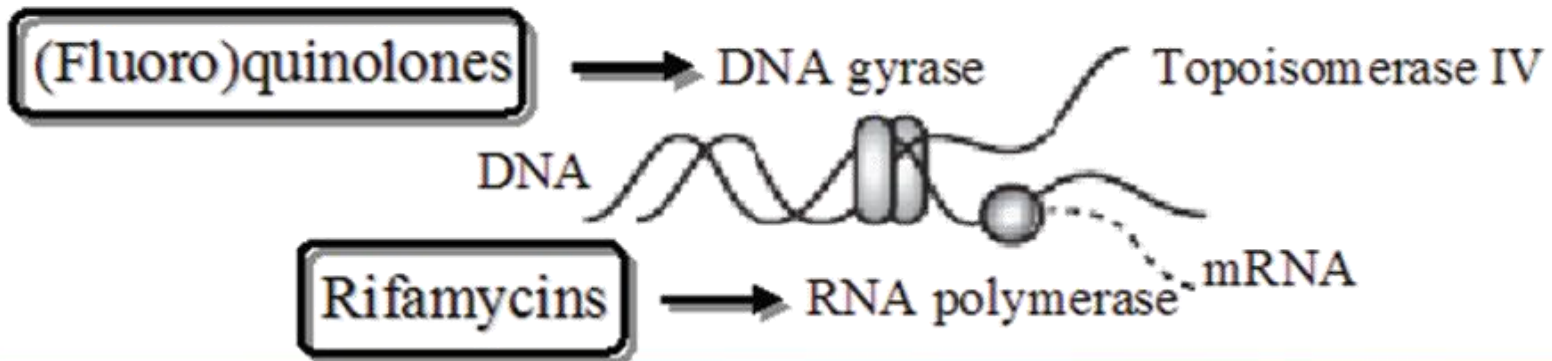
抑制核酸合成

- 抑制DNA合成(抑制DNA gyrase)

Quinolones: nalidixic acid, ofloxacin, ciprofloxacin, levofloxacin, moxifloxacin, gemifloxacin

- 抑制RNA合成：Rifampicin

DNA/RNA biosynthesis





抑制細胞膜功能

- Polymyxins: Polycation molecule binds to membrane and change membranous electric charges
- Daptomycin: Insert phosphatidyl glycerol and depolarization



抑制營養代謝及生長

- Sulfonamides: inhibit folic acid synthesis

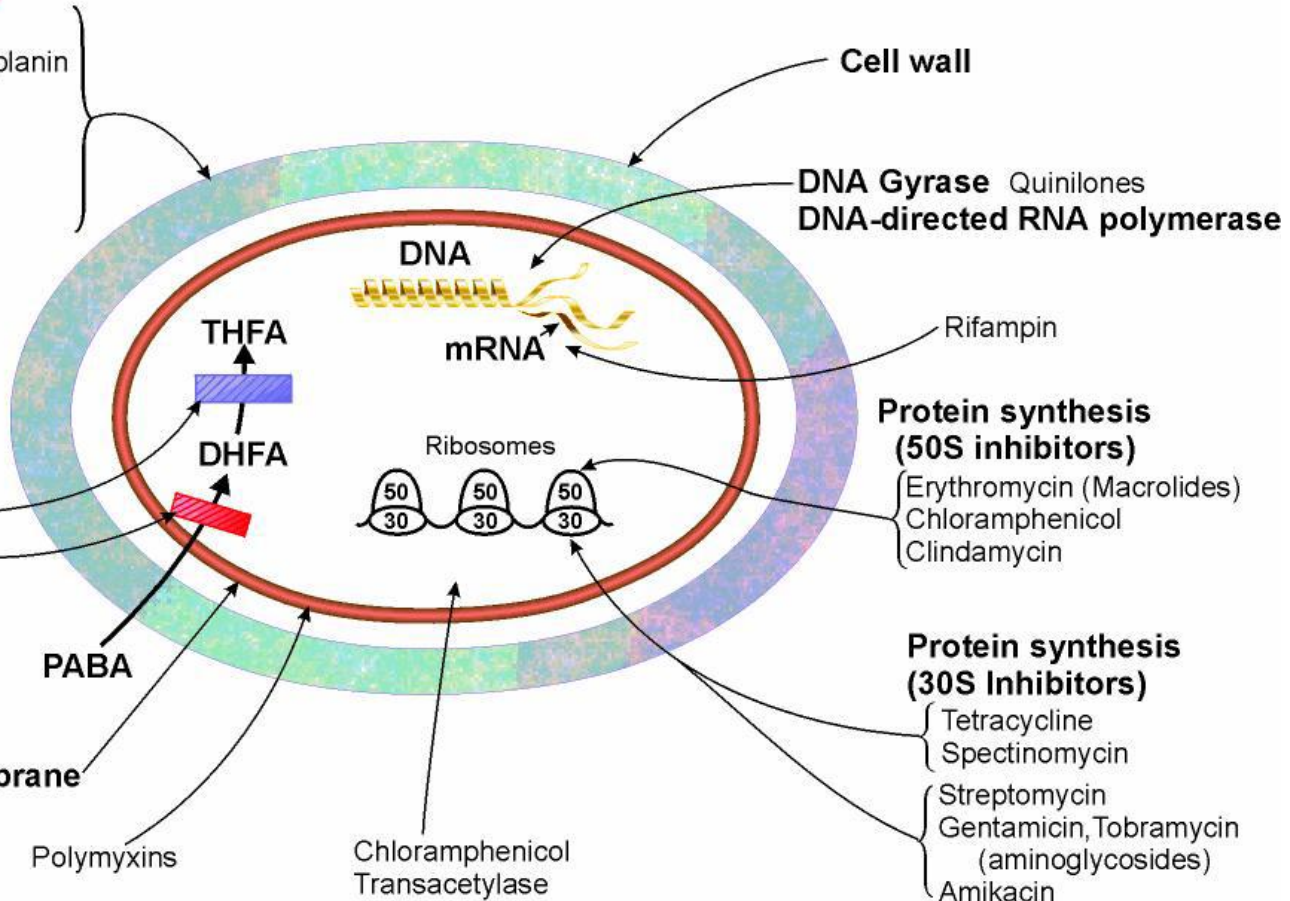
Bacterial Targets for Current Antibiotics Used in the Clinic

Cell wall synthesis

Cycloserine
Vancomycin, Teichoplanin
Bacitracin
Penicillins
Cephalosporins
Monobactams
Carbapenems

Folic acid metabolism

Trimethoprim
Sulfonamides



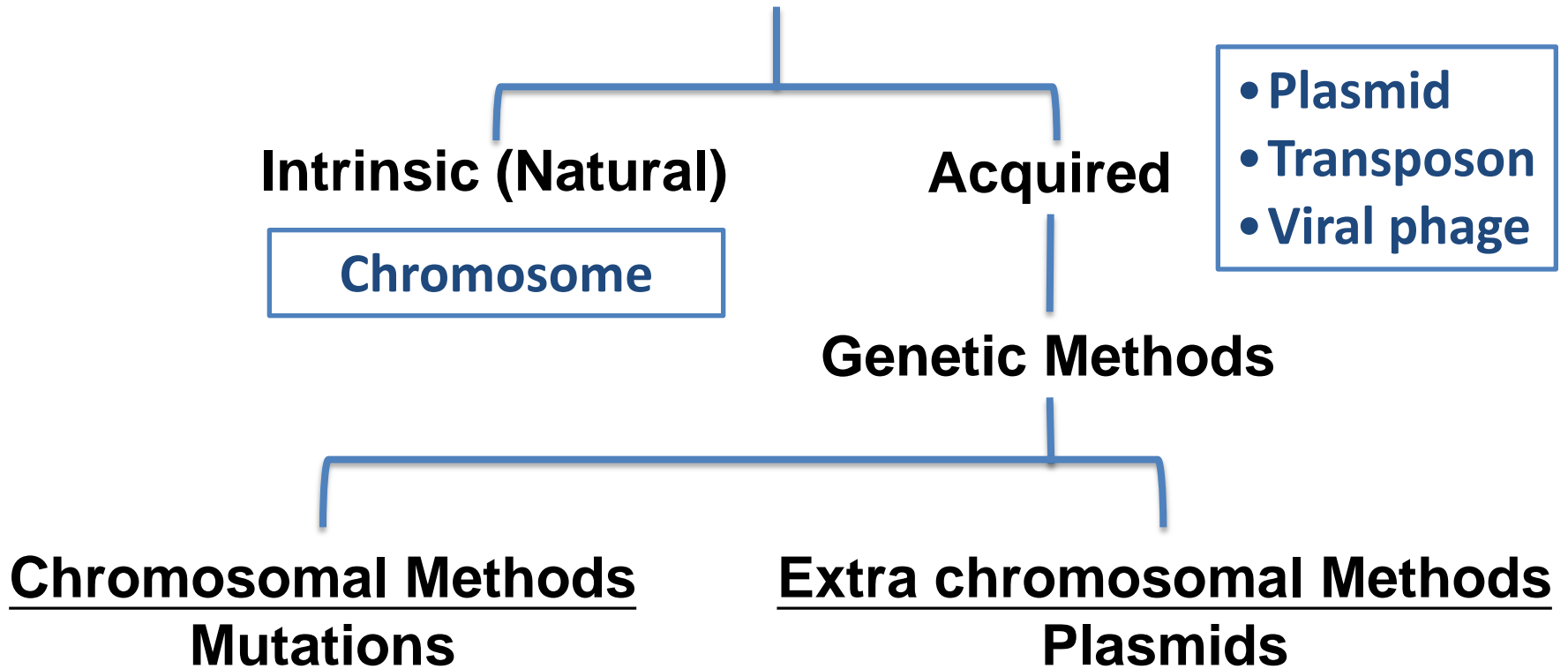
資料來源：Dyner, L. (2009). Antibiotic deployment and resistance observed. [Chart from Slideshow]. Stanford University. Retrieved from Slideshow from <http://peds.stanford.edu/Tools/documents/AntibioticResistanceLLD.pdf>



2. 細菌抗藥性機轉之演進



Mechanism of Antibiotic Resistance





Intrinsic/Acquired Resistance

Example: *Enterococcus*

Intrinsic resistance

β -Lactams (particularly cephalosporins and penicillinase-resistant penicillins)

Low concentrations of aminoglycosides

Clindamycin

Fluoroquinolones

Trimethoprim-sulfamethoxazole

Acquired resistance

High concentrations of β -lactams, through penicillin-binding proteins or β -lactamase

High concentrations of aminoglycosides

Glycopeptides (vancomycin and teicoplanin)

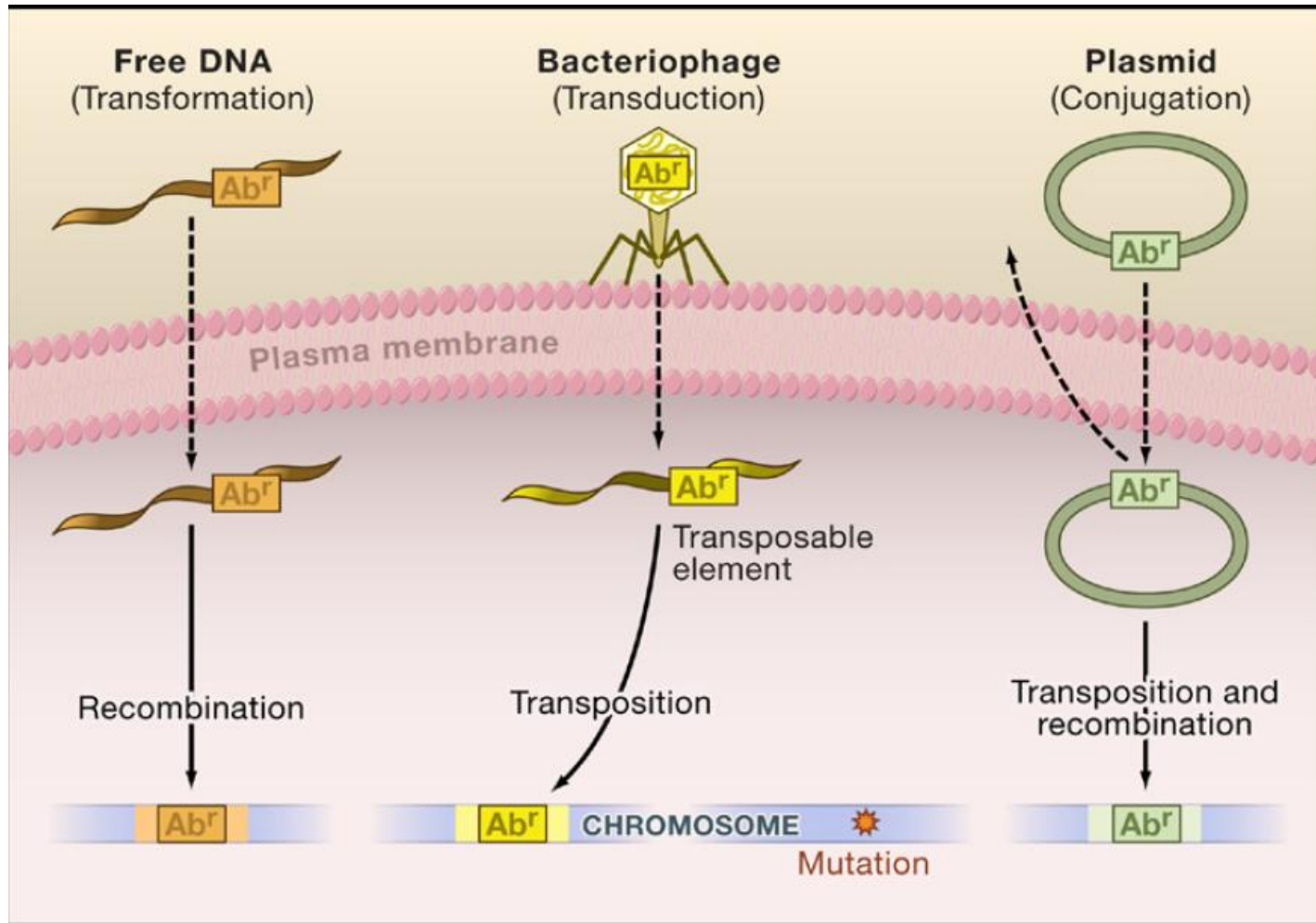
Tetracycline

Erythromycin

Fluoroquinolones

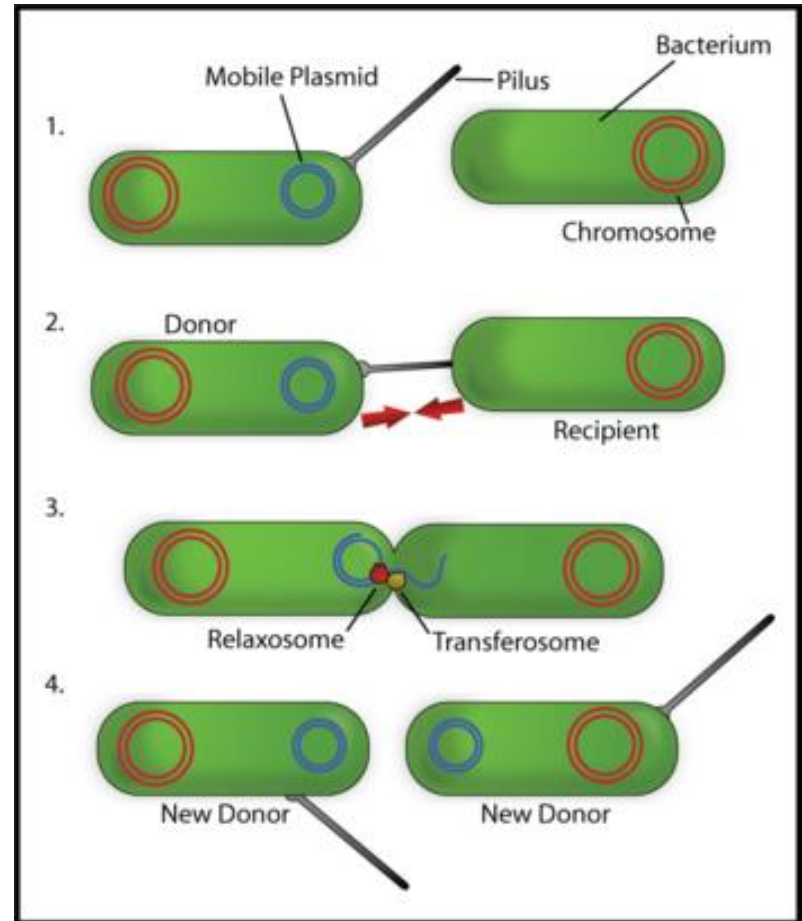


Acquisition of Antibiotic Resistance



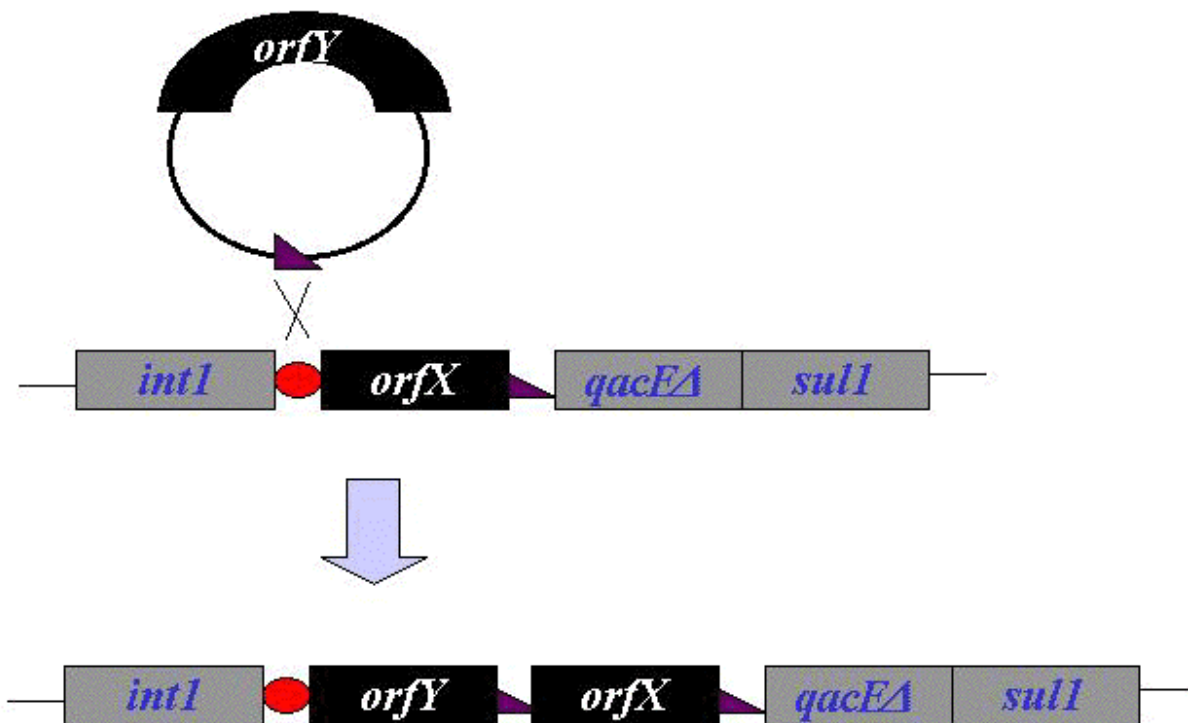
Horizontal Spreads of Antibiotic Resistance Genes(1/2)

- Plasmid
- Transposon

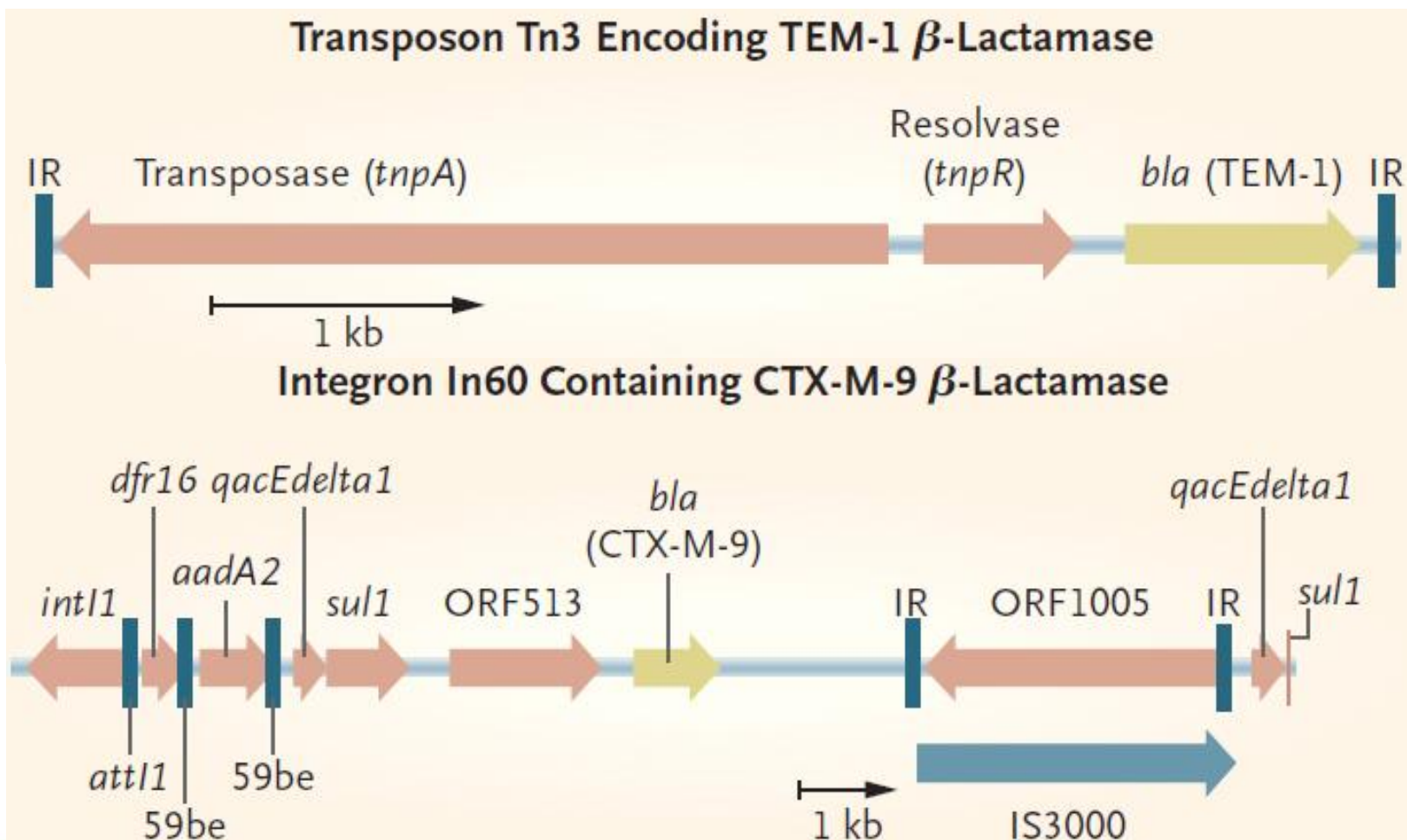


Horizontal Spreads of Antibiotic Resistance Genes(2/2)

- Gene cassette
- Integron



Transposons and Integrons



細菌發生抗藥性的步驟

抗藥性基因之發生

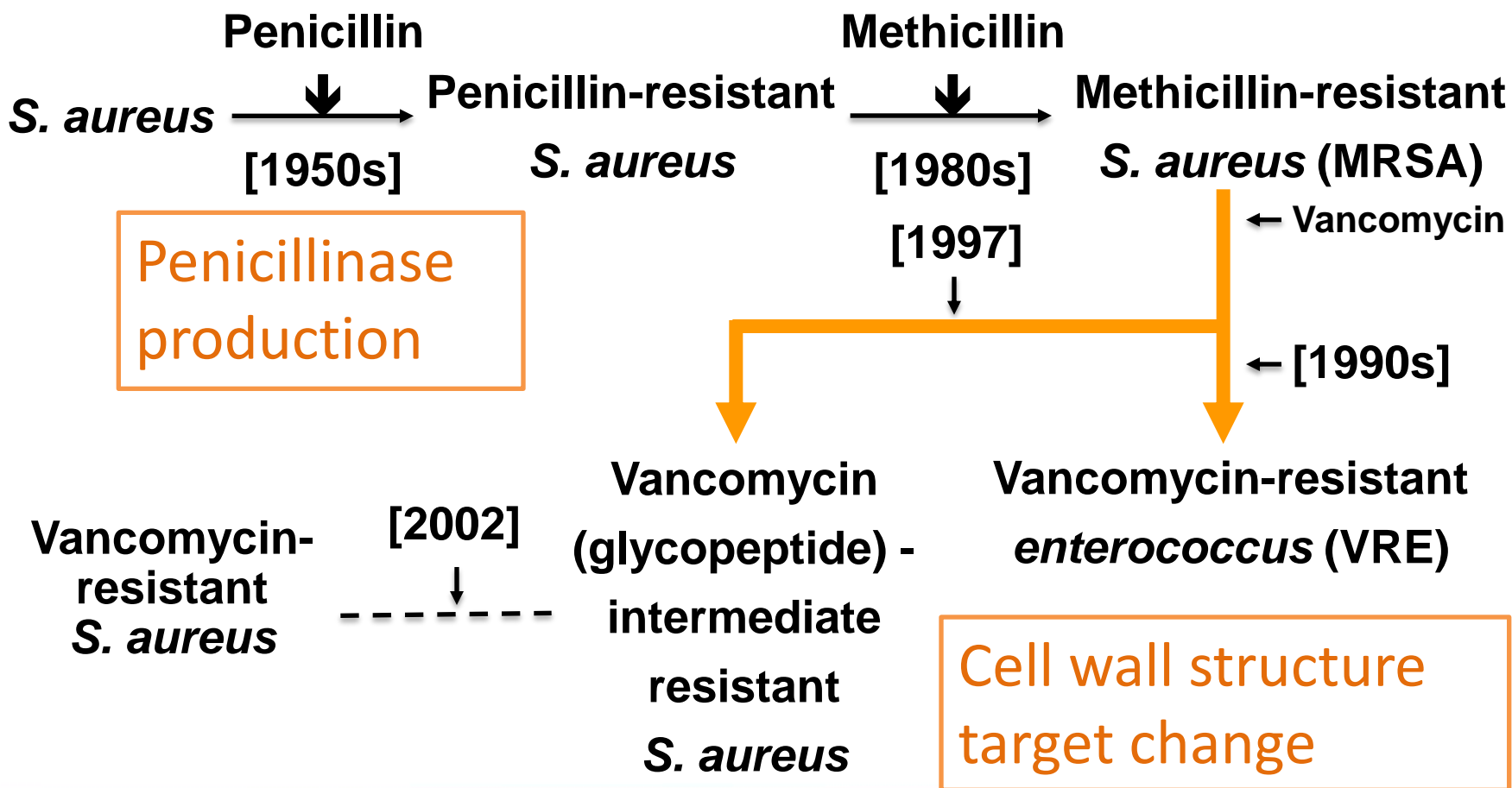
(基因本身自然突變；自其他生物獲得)

環境選擇性壓力 (Selection pressure)
(Antibiotics use in human and animal)

抗藥性細菌之散佈

(Spread in the environment)
(未做好院內感控)

抗藥性機轉的演進 以金黃色葡萄球菌為例





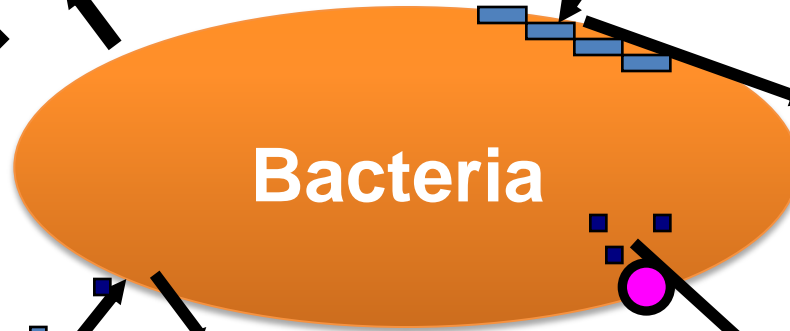
Mechanisms of Antibiotic Resistance

- Enzymatic alteration
- Decreased permeability
- Efflux
- Alteration/protection of target site
- Overproduction of target
- Bypass of inhibited process
- Bind-up antibiotic

抗藥性細菌的抗藥機轉

Produce enzyme to inactivate drugs

Modify binding targets



Drug penetration barrier (porins)

Drug efflux pumping



Resistant Mechanism of Common Pathogens

ANTIBIOTIC CLASS

MECHANISM OF RESISTANCE

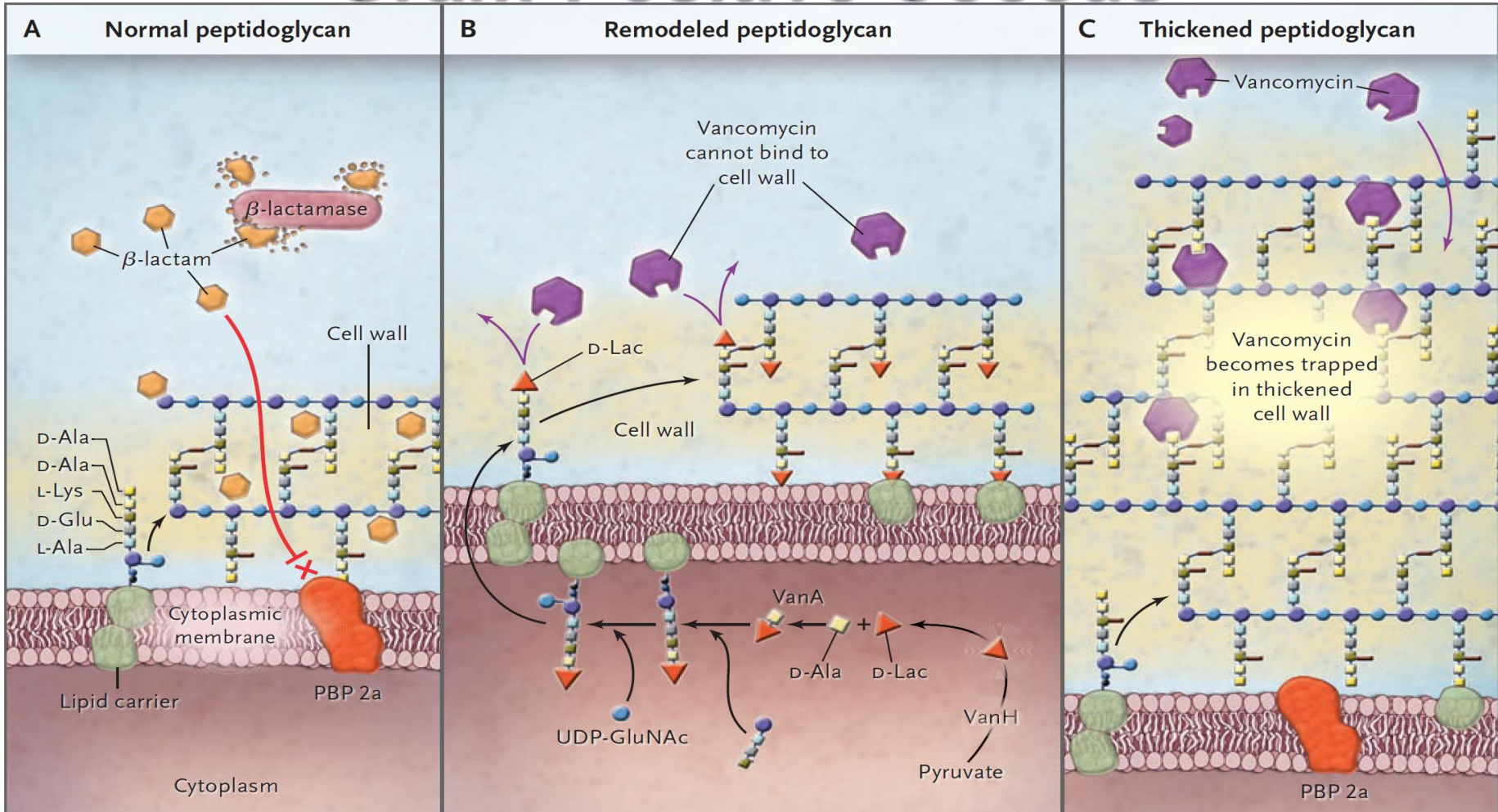
Cephalosporins	Extended-spectrum β -lactamases, chromosomal cephalosporinases
β -Lactamase inhibitors	Hyperproducers of β -lactamases, new β -lactamases resistant to inhibitors, chromosomal cephalosporinases
Carbapenems	Zinc metalloenzymes and other β -lactamases
Vancomycin, teicoplanin	Modified cell-wall precursors with decreased affinity for vancomycin
Quinolones	Alterations in DNA topoisomerase, efflux mechanisms, permeability changes
Trimethoprim-sulfamethoxazole	Resistant enzymes in folate-synthesis pathway
Erythromycin, new macrolides	Methylation of the bacterial ribosome producing resistance to macrolides, clindamycin, and streptogramin B antibiotics
Aminoglycosides	Aminoglycoside-modifying enzymes



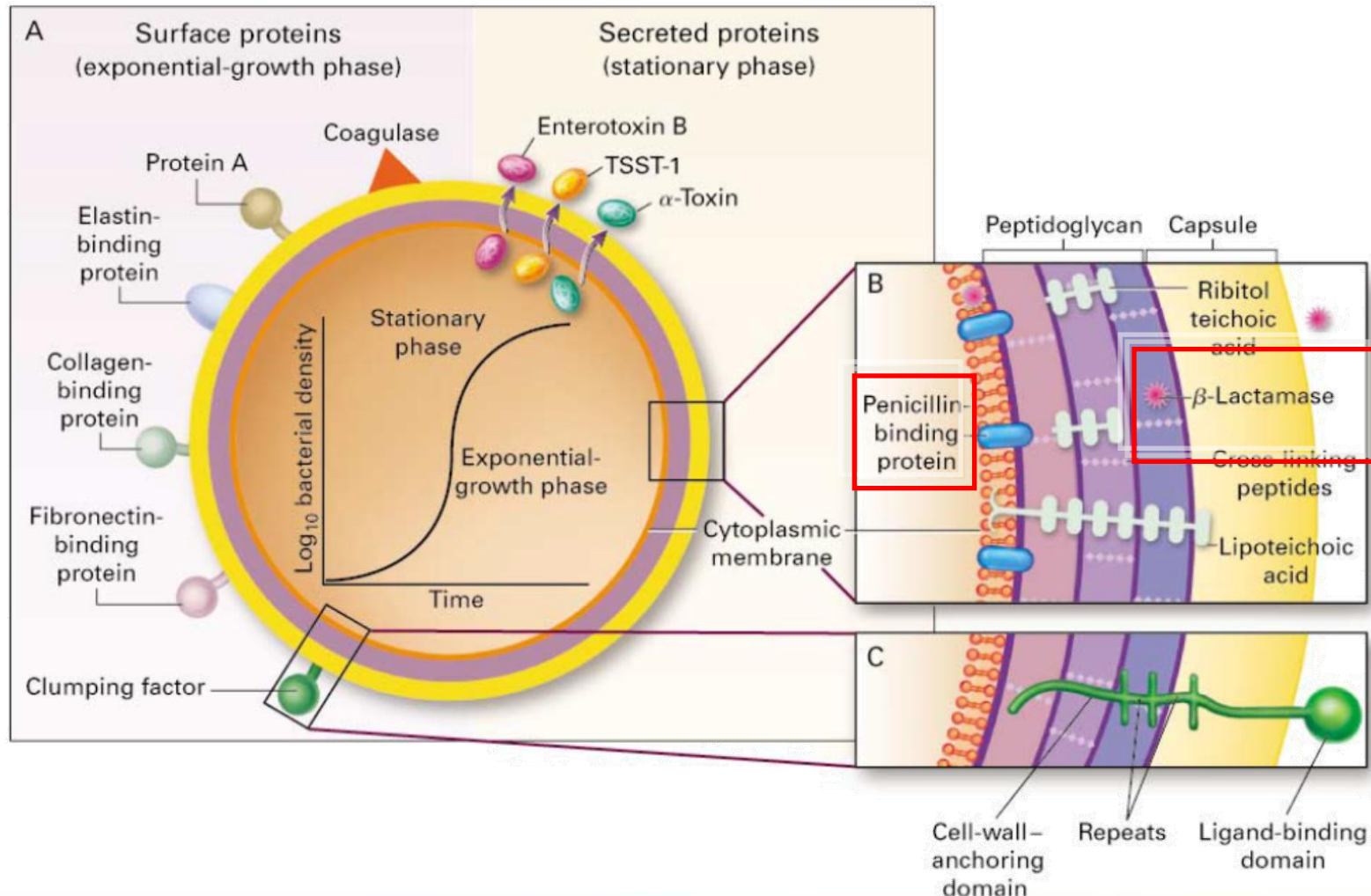


3. 抗藥性革蘭氏陽性菌之抗藥機轉

Resistant Mechanism in Gram-Positive Coccus



Structure of *Staphylococcus aureus*





Methicillin-resistant *Staphylococcus aureus* (MRSA)(1/2)

- Resistant to methicillin also resistant to most penicillins and cephalosporins
- High level methicillin resistance by *mec* gene that encodes penicillin-binding protein 2a
- Horizontal transfer of *mec* DNA
- The expression of resistance to methicillin is often heterogeneous



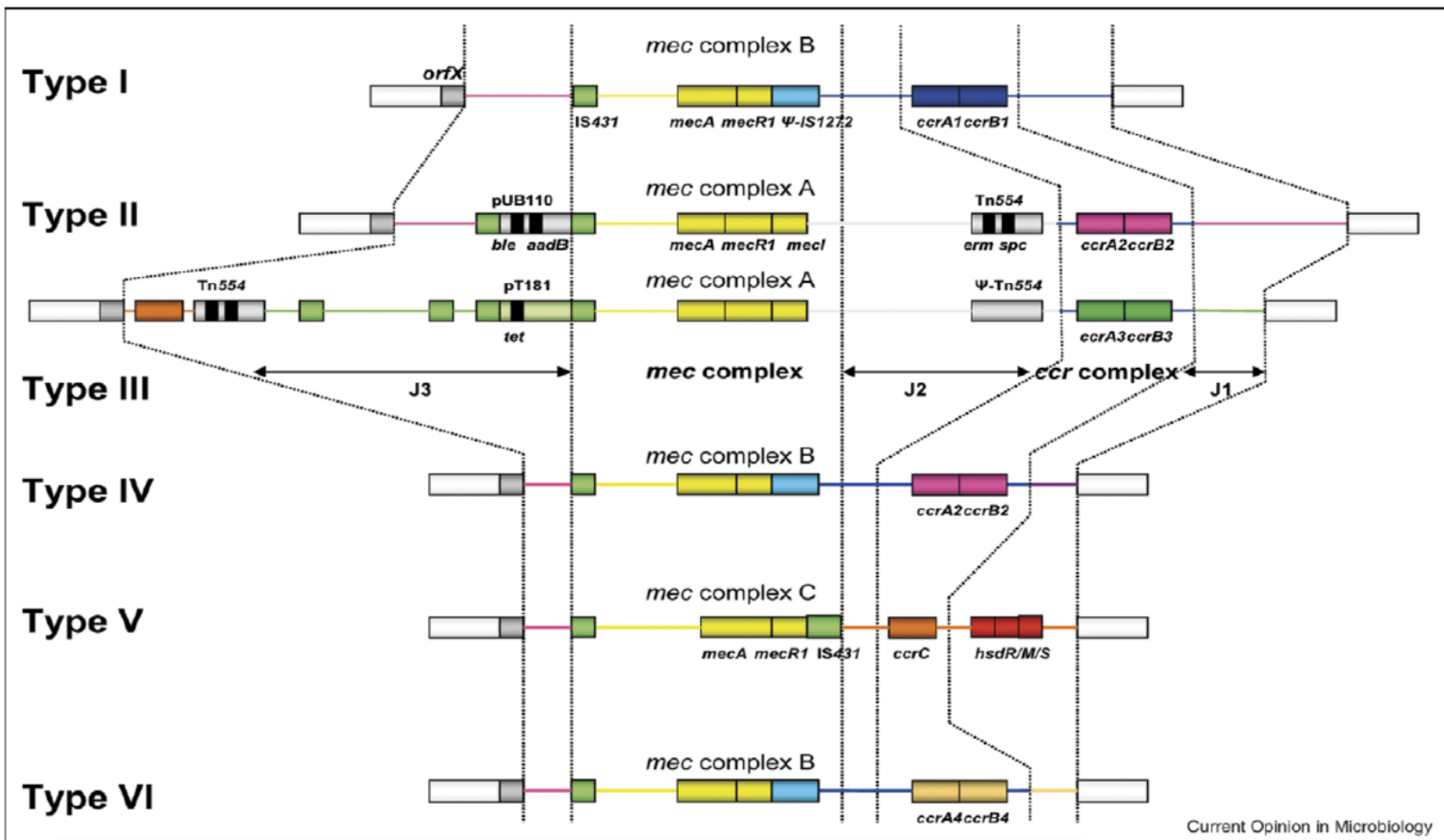


Methicillin-resistant *Staphylococcus aureus* (MRSA)(2/2)

- Reduced-affinity penicillin-binding protein (PBP2a or PBP2').
- Resistant to semi-synthetic penicillin and other β -lactams.
- PBP2a encoded by *mecA* gene, resides within a genomic island, staphylococcal cassette chromosome *mec* (SCC*mec*).
- At least 6 SCC types (I-VI) identified.



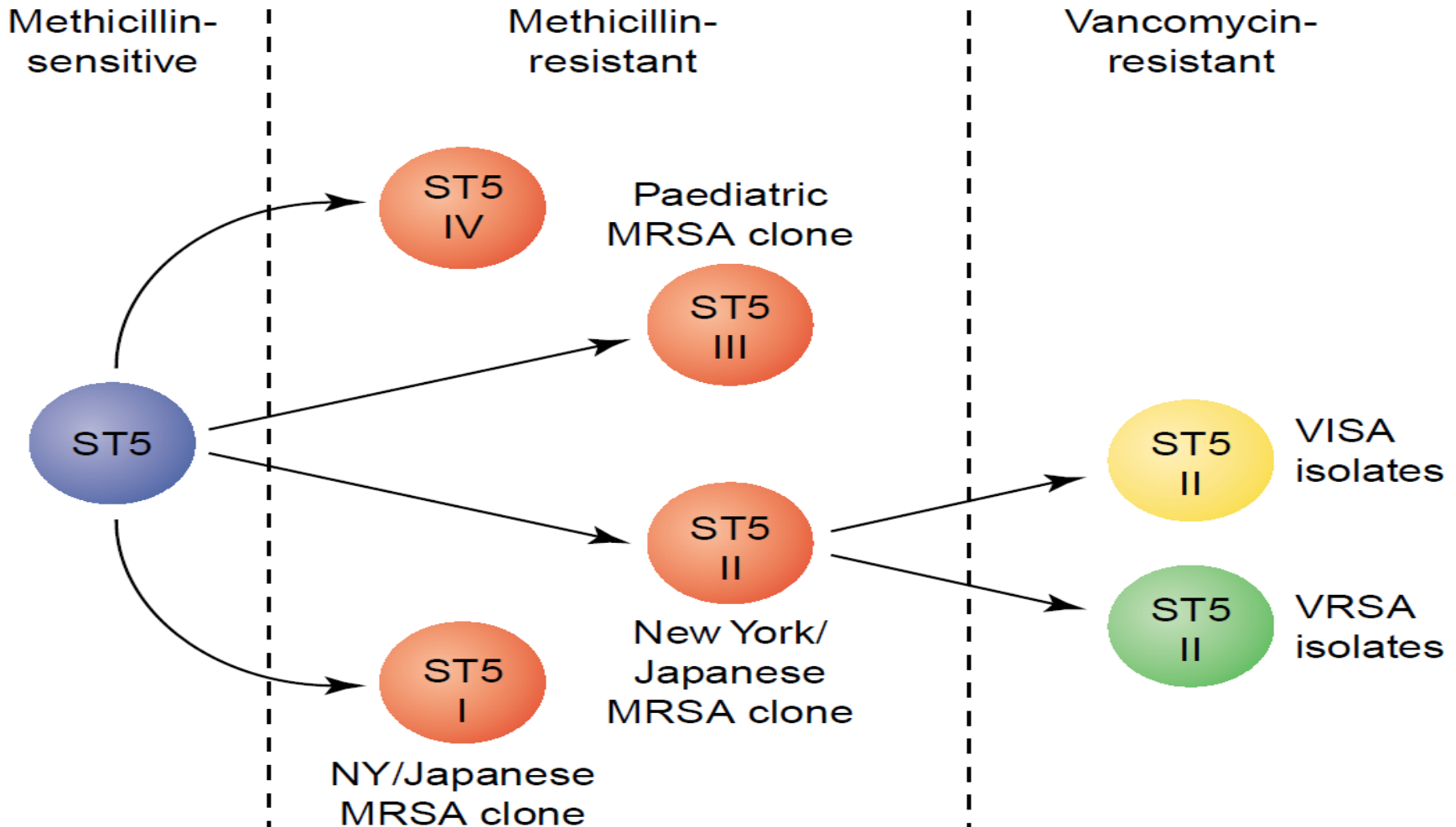
MRSA, SCCmec Types I–VI



Current Opinion in Microbiology

資料來源：De Lencastre H, et al. Current Opinion in Microbiology 2007; 10:428–435

Evolution of Increasing Resistance in ST5





Vancomycin-resistant *Enterococcus* (VRE)

- Van A strains
 - Teicoplanin (MICs 16-512 mg/l)
 - Vancomycin (MICs 64-1000 mg/l)
- Van B strains
 - Teicoplanin (MICs 0.25-2 mg/l)
 - Vancomycin (MICs 4-1000 mg/l)
- Van C strains
 - Teicoplanin (MICs 0.12-2 mg/l)
 - Vancomycin (MICs 2-32 mg/l)





Glycopeptide-resistant *Enterococcus*

CHARACTERISTIC	PHENOTYPE		
	VANA	VANB	VANC
Vancomycin MIC ($\mu\text{g/ml}$)	64 to >1000	4 to 1024	2 to 32
Teicoplanin MIC ($\mu\text{g/ml}$)	16 to 512	≤ 0.5	≤ 0.5
Most frequent enterococcal species	<i>Ent. faecium</i> <i>Ent. faecalis</i>	<i>Ent. faecalis</i> <i>Ent. faecium</i>	<i>Ent. gallinarum</i> <i>Ent. casseliflavus</i>
Genetic determinant	Acquired	Acquired	Intrinsic
Transferable	Yes	Yes	No



Site of Action of Glycopeptides

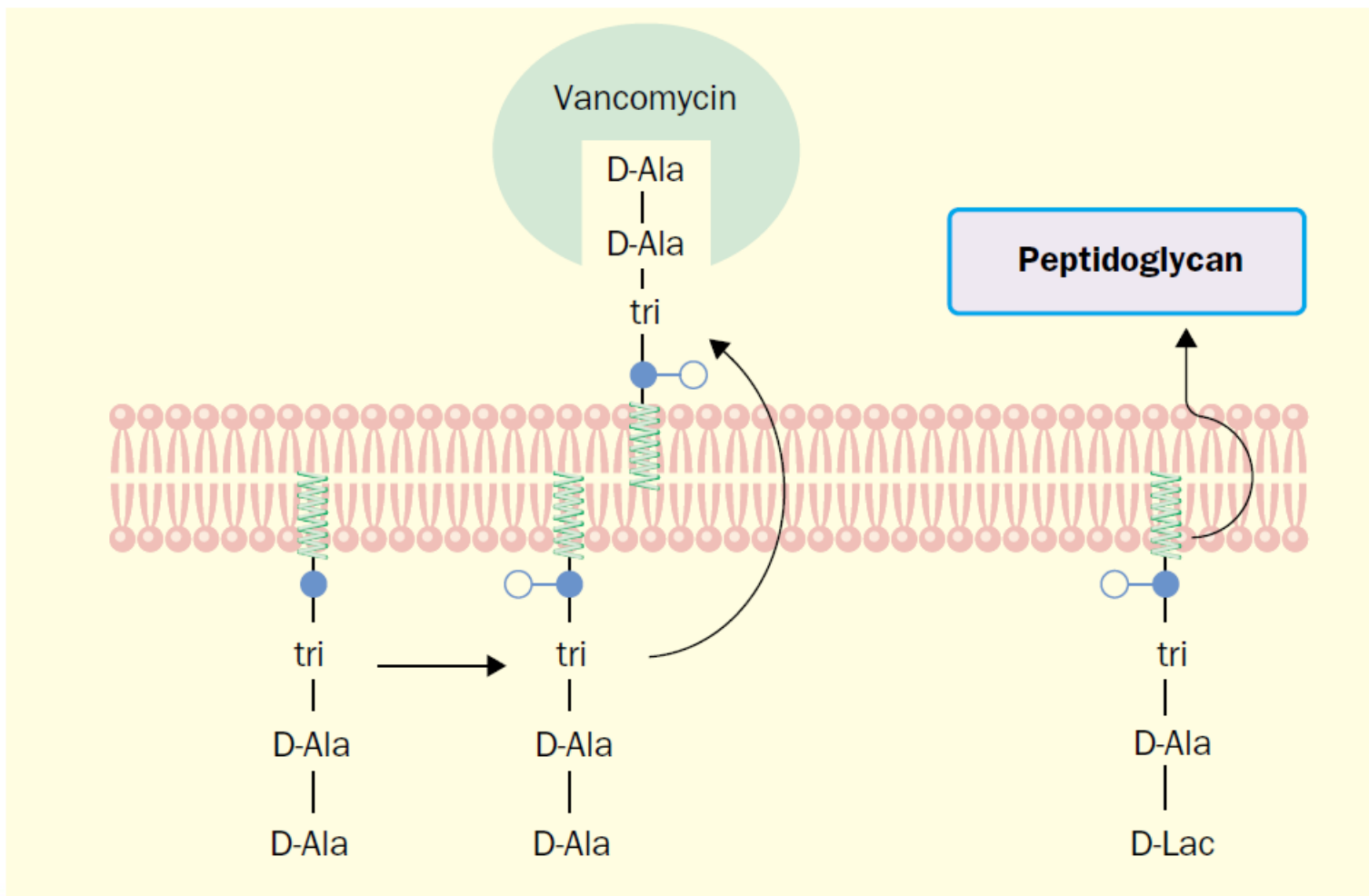
Pentapeptide



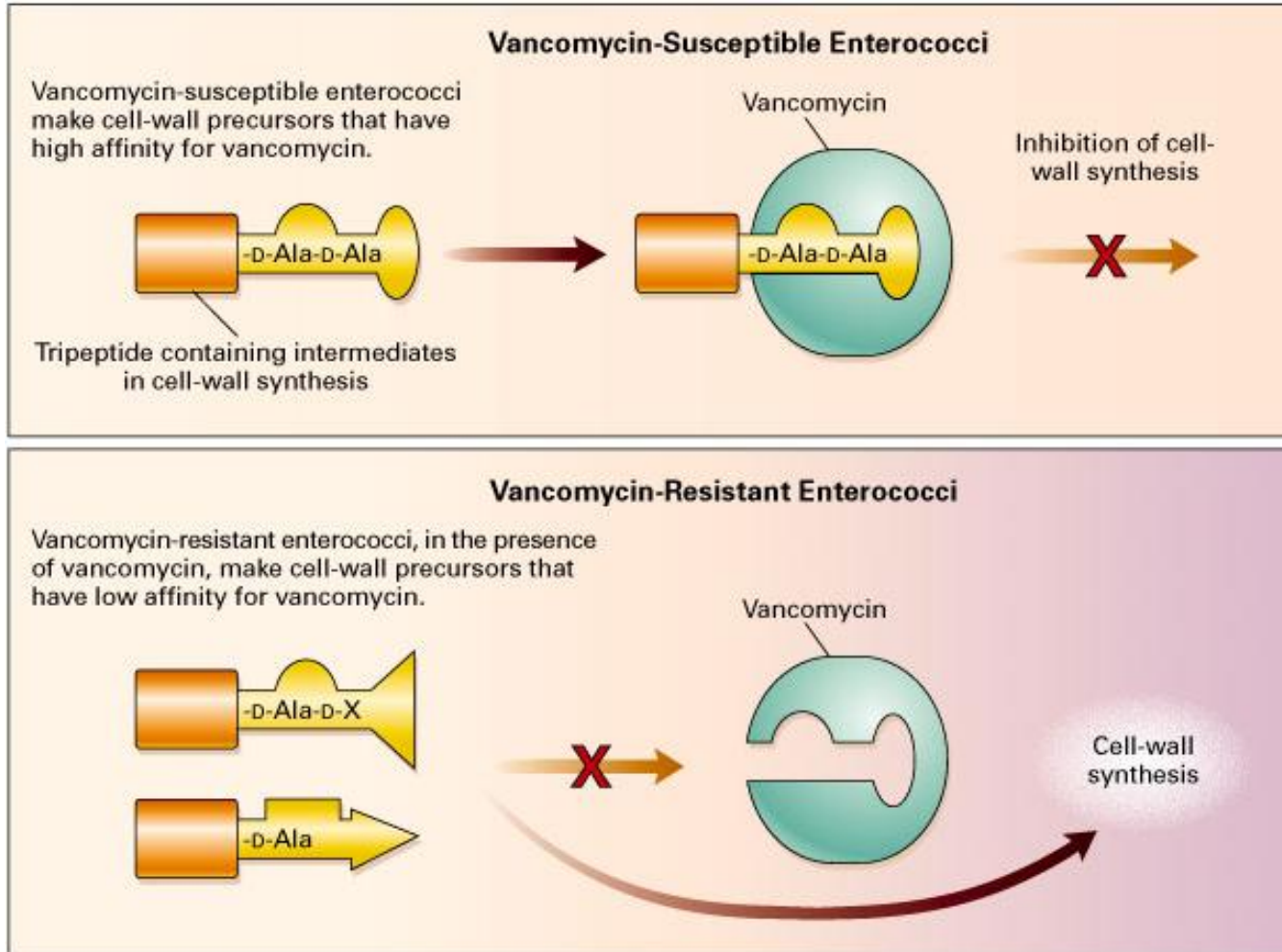
○ = N-Acetyl glucosamine-N-Acetyl muramic acid
aa = amino acid , D-ala = D-alanine

★ **Binds to terminal D-ala-D-ala dipeptide prevent pentapeptide being used for cell wall synthesis**

VanA-Type Resistance in *Enterococcus*

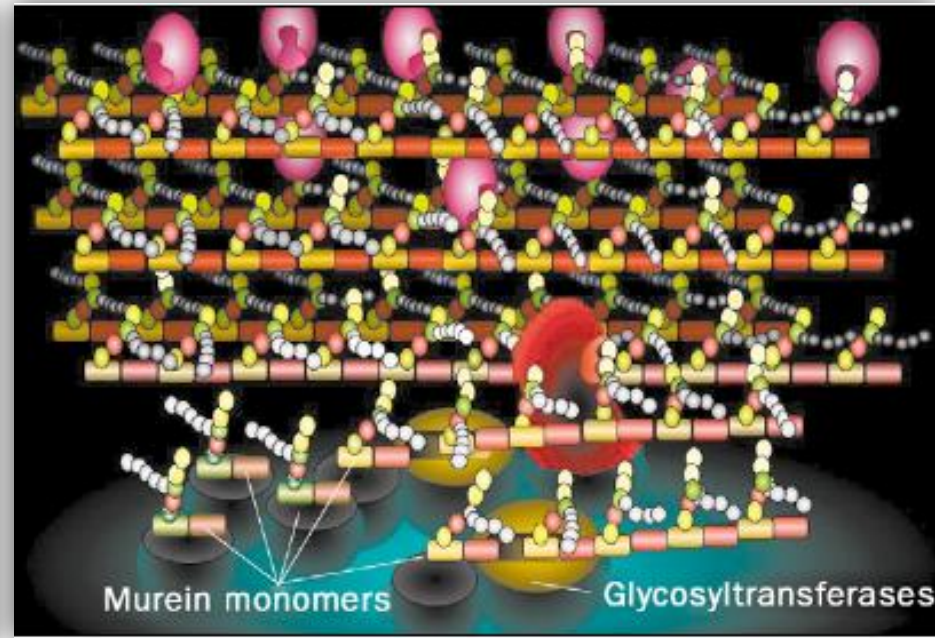
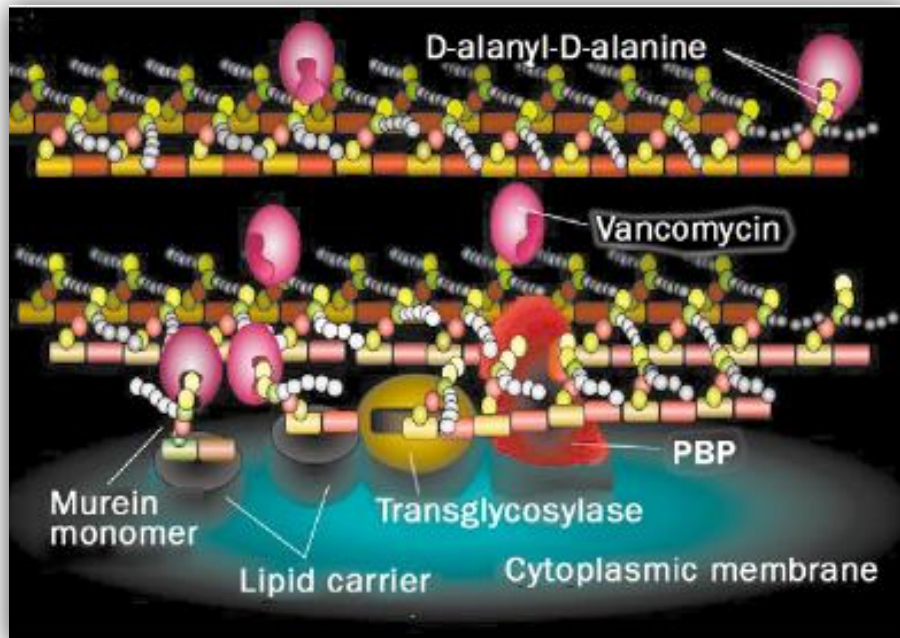


Mechanism of Vancomycin Resistance in *Enterococcus*



Resistant Mechanism for VRSA

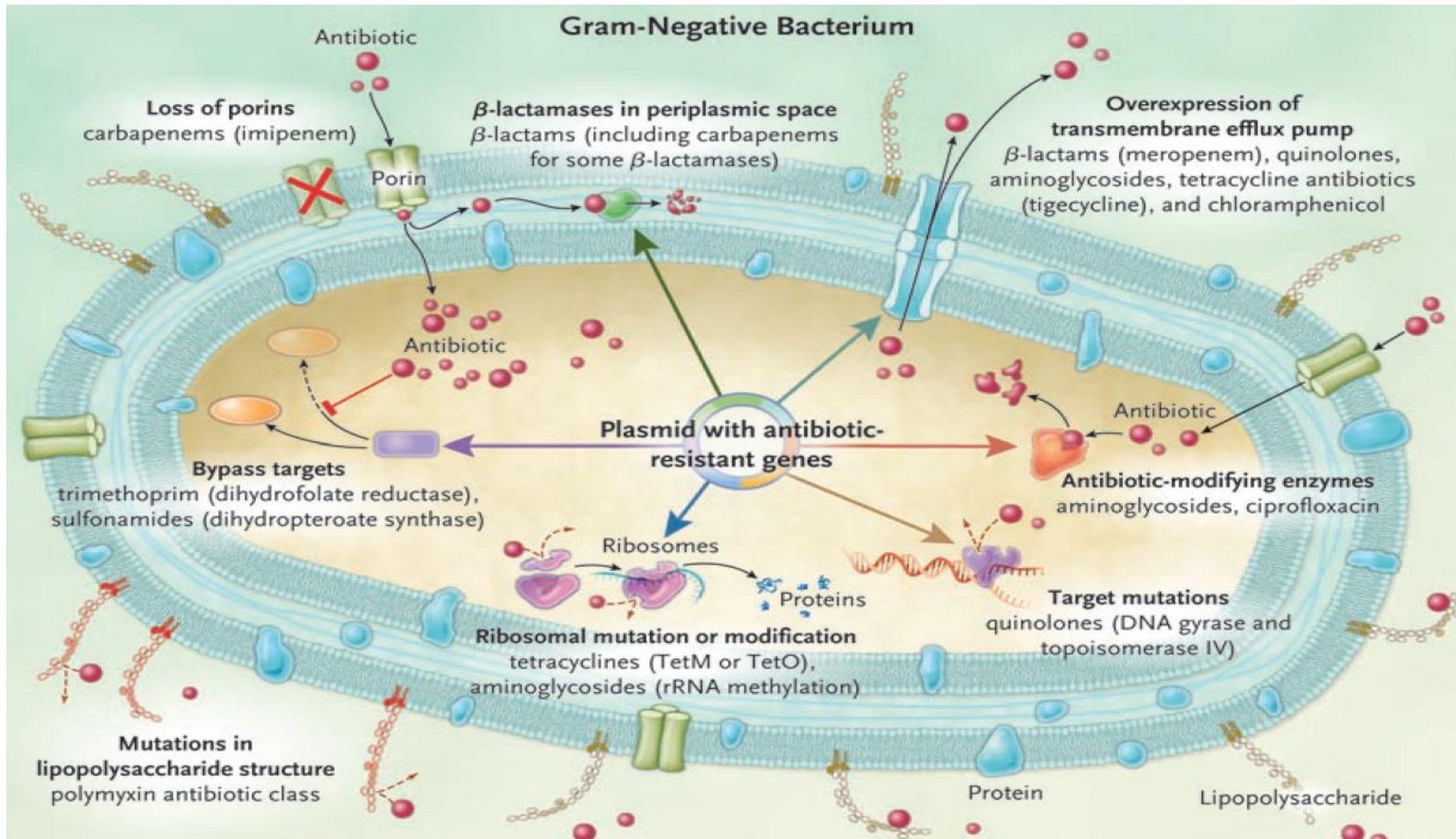
- Vancomycin resistance is acquired by mutation and thickening of cell wall due to accumulation of excess amounts of peptidoglycan.





4. 抗藥性革蘭氏陰性菌之抗藥機轉

Resistant Mechanisms of Gram-Negative Bacterium





Major Resistant Gram-Negative Bacteria: β -Lactamase

Drug-Resistant Pathogen

Extended-spectrum β -lactamase-producing
Enterobacteriaceae

Carbapenemase-producing
Enterobacteriaceae

Carbapenem-resistant *Pseudomonas*
aeruginosa and *Acinetobacter baumannii*





Resistance in Enterobacteriaceae

β -Lactamase	Examples	Substrates	Molecular class
Broad-spectrum	TEM-1, TEM-2, SHV-1	Penicillin G, aminopenicillins, carboxypenicillins, piperacillin, narrow-spectrum cephalosporins	A
	OXA family	Substrates of the broad-spectrum group plus cloxacillin, methicillin, and oxacillin	D
Extended-spectrum	TEM family, SHV family	Substrates of the broad-spectrum group plus oxyimino-cephalosporins, and monobactam (aztreonam)	A
	CTX-M family	Substrates of the expanded-spectrum group plus, for some enzymes, cefepime	A
	OXA family	Same as for CTX-M family	D
	Others (PER-1, PER-2, BES-1, GES/IBC family, SFO-1, TLA-1, VEB-1, VEB-2)	Same as for TEM family and SHV family	A
AmpC	ACC-1, ACT-1, CFE-1, CMY family, DHA-2, FOX family, LAT family, MIR-1, MOX-1, MOX-2	Substrates of expanded-spectrum group plus cephamycins	C
Carbapenemase	IMP family, VIM family, GIM-1, SPM-1 (metallo- β -enzymes)	Substrates of the expanded-spectrum group plus cephamycins and carbapenems	B
	KPC-1, KPC-2, KPC-3	Same as for IMP family, VIM family, GIM-1, and SPM-1	A
	OXA-23, OXA-24, OXA-25, OXA-26, OXA-27, OXA-40, OXA-48	Same as for IMP family, VIM family, GIM-1, and SPM-1	D





Ambler Molecular Classification of β -Lactamases(1/2)

β -lactamase-class	β -lactamases	Important examples	Preferential occurrence	Important phenotypical resistance traits ^a
A Serine	Broad-spectrum β -lactamases	TEM-1, TEM-2 SHV-1, SHV-11	Enterobacteriaceae and nonfermenters ^b	ampicillin, cephalotin
	ESBL TEM-type	TEM-3, TEM-52		penicillins, 3rd gen. cephalosporins
	ESBL SHV-type	SHV-5, SHV-12		
	ESBL CTX-M-type	CTX-M-1, CTX-M-15		
	Carbapenemases	KPC, GES, SME		all β -Lactams ^c
C Serine	AmpC cephamycinases (chromosomal-encoded)	AmpC	Enterobacter spp. Citrobacter spp.	cephamycins (cefotaxime), 3rd gen. cephalosporins





Ambler Molecular Classification of β -Lactamases(2/2)

β -lactamase-class	β -lactamases	Important examples	Preferential occurrence	Important phenotypical resistance traits ^a
D Serine	AmpC cephamycinases (plasmid-encoded)	CMY, DHA, MOX FOX, ACC,	Enterobacteriaceae	cephamycins (cefoxitin), 3rd gen. cephalosporins
	Broad-spectrum β -lactamases	OXA-1, OXA-9	Enterobacteriaceae; <i>A. baumannii</i>	oxacillin, ampicillin cephalotin
	ESBL OXA-type	OXA-2, OXA-10		penicillins, 3rd gen. cephalosporins
	Carbapenemases; Carbapenemases	OXA-48; OXA-23,-24,-58		ampicillin, imipenem; all β -lactams ^c
B MBL-Zn	Metallo- β -lactamases (Carbapenemases)	VIM IMP	Enterobacteriaceae and nonfermenters	all β -lactams ^c





Bush-Jacoby Functional Classification of β -Lactamases(1/2)

Bush-Jacoby group (2009)	Bush-Jacoby-Medeiros group (1995)	Molecular class (subclass)	Distinctive substrate(s)	Inhibited by		Defining characteristic(s)	Representative enzyme(s)
				CA or TZB ^a	EDTA		
1	1	C	Cephalosporins	No	No	Greater hydrolysis of cephalosporins than benzylpenicillin; hydrolyzes cephamycins	<i>E. coli</i> AmpC, P99, ACT-1, CMY-2, FOX-1, MIR-1
1e	NI ^b	C	Cephalosporins	No	No	Increased hydrolysis of ceftazidime and often other oxyimino- β -lactams	GC1, CMY-37
2a	2a	A	Penicillins	Yes	No	Greater hydrolysis of benzylpenicillin than cephalosporins	PC1
2b	2b	A	Penicillins, early cephalosporins	Yes	No	Similar hydrolysis of benzylpenicillin and cephalosporins	TEM-1, TEM-2, SHV-1
2be	2be	A	Extended-spectrum cephalosporins, monobactams	Yes	No	Increased hydrolysis of oxyimino- β -lactams (cefotaxime, ceftazidime, ceftriaxone, cefepime, aztreonam)	TEM-3, SHV-2, CTX-M-15, PER-1, VEB-1
2br	2br	A	Penicillins	No	No	Resistance to clavulanic acid, sulbactam, and tazobactam	TEM-30, SHV-10
2ber	NI	A	Extended-spectrum cephalosporins, monobactams	No	No	Increased hydrolysis of oxyimino- β -lactams combined with resistance to clavulanic acid, sulbactam, and tazobactam	TEM-50
2c	2c	A	Carbenicillin	Yes	No	Increased hydrolysis of carbenicillin	PSE-1, CARB-3
2ce	NI	A	Carbenicillin, cefepime	Yes	No	Increased hydrolysis of carbenicillin, cefepime, and ceftipime	RTG-4
2d	2d	D	Cloxacillin	Variable	No	Increased hydrolysis of cloxacillin or oxacillin	OXA-1, OXA-10



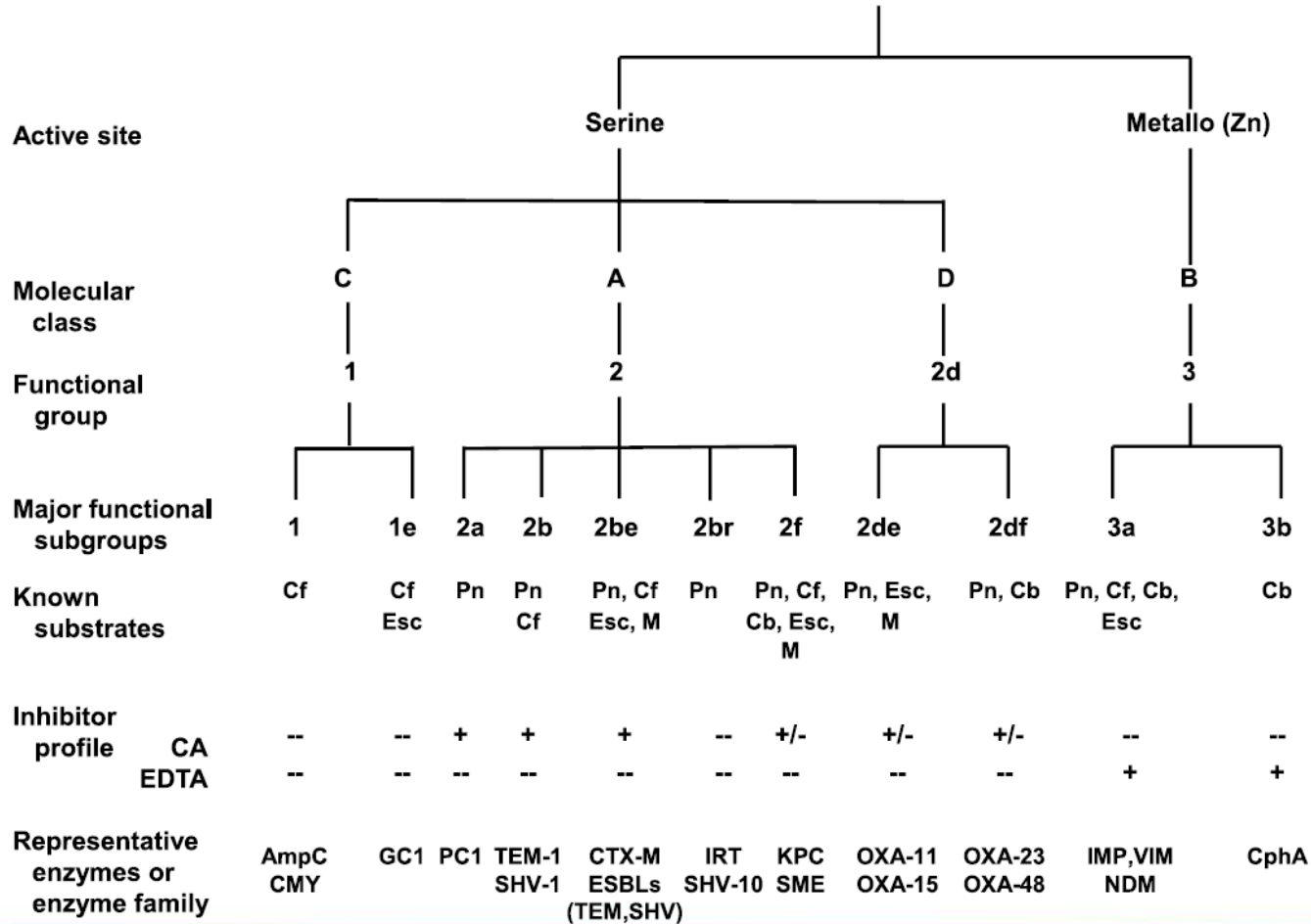


Bush-Jacoby Functional Classification of β -Lactamases(2/2)

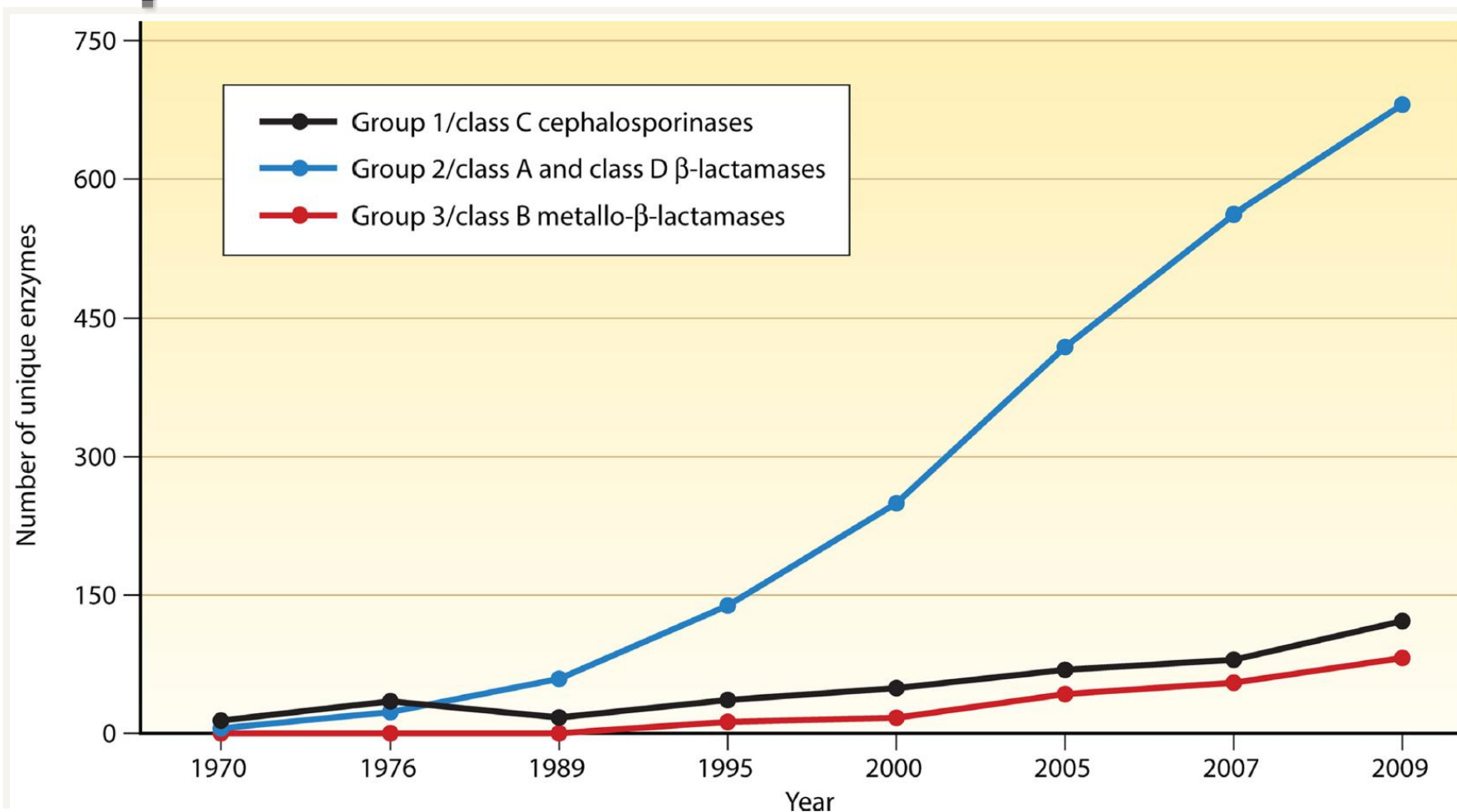
Bush-Jacoby group (2009)	Bush-Jacoby-Medeiros group (1995)	Molecular class (subclass)	Distinctive substrate(s)	Inhibited by		Defining characteristic(s)	Representative enzyme(s)
				CA or TZB ^a	EDTA		
2de	NI	D	Extended-spectrum cephalosporins	Variable	No	Hydrolyzes cloxacillin or oxacillin and oxyimino- β -lactams	OXA-11, OXA-15
2df	NI	D	Carbapenems	Variable	No	Hydrolyzes cloxacillin or oxacillin and carbapenems	OXA-23, OXA-48
2e	2e	A	Extended-spectrum cephalosporins	Yes	No	Hydrolyzes cephalosporins. Inhibited by clavulanic acid but not aztreonam	CepA
2f	2f	A	Carbapenems	Variable	No	Increased hydrolysis of carbapenems, oxyimino- β -lactams, cephamycins	KPC-2, IMI-1, SME-1
3a	3	B (B1)	Carbapenems	No	Yes	Broad-spectrum hydrolysis including carbapenems but not monobactams	IMP-1, VIM-1, CcrA, IND-1



Molecular/Functional Features of Major β -Lactamases




Increasing No. of Group 1, 2, and 3 β -Lactamases from 1970 to 2009





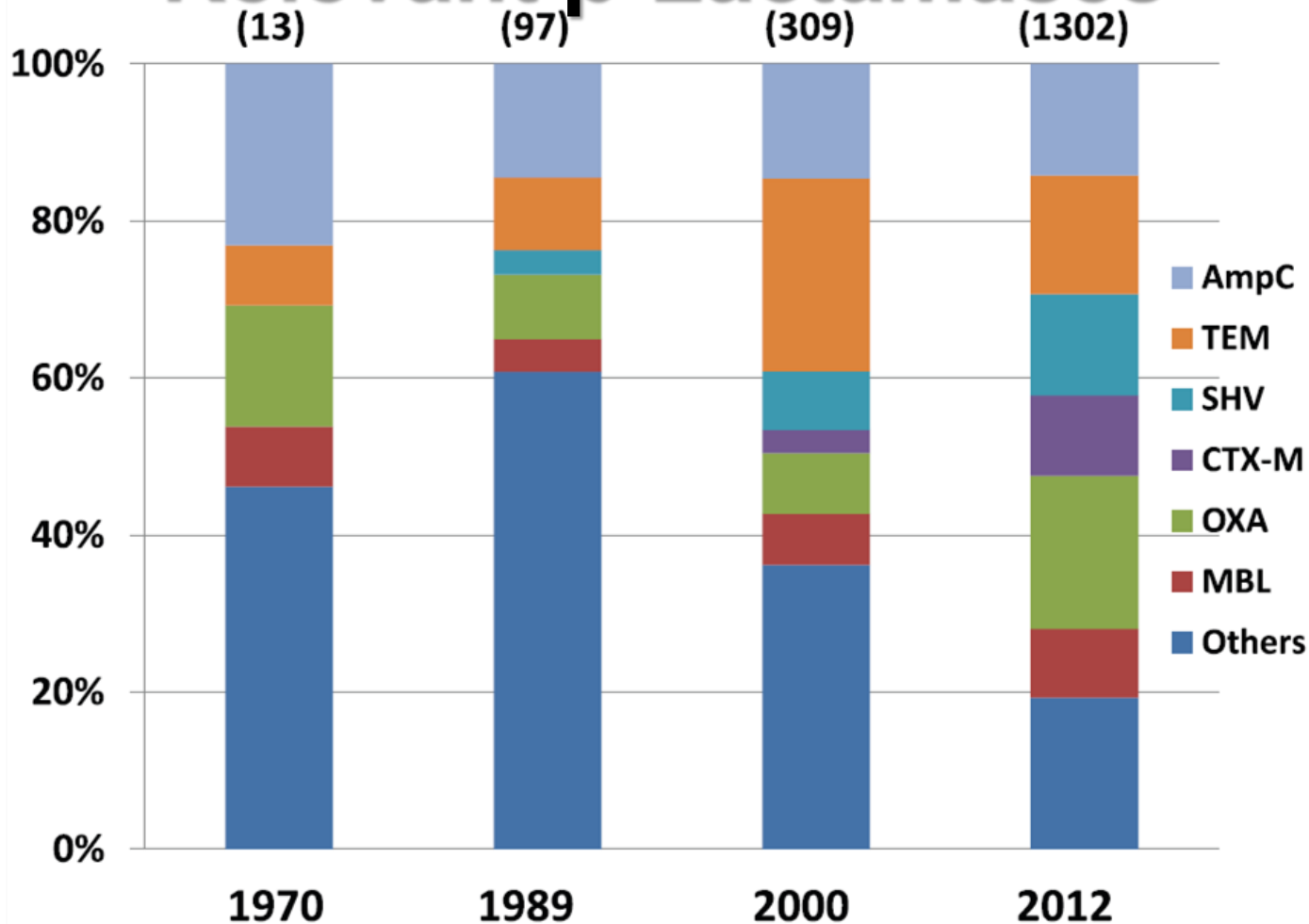
Extended-Spectrum β -Lactamase-Producing Enterobacteriaceae

	Country
CTX-M1	Italy
CTX-M2	Israel, Argentina
CTX-M3	Poland
CTX-M9	Spain
CTX-M14	Spain, Canada, China
CTX-M15	Worldwide

 Most common CTX-M enzymes



Significance of Clinically Relevant β -Lactamases





Selected β -Lactamases of Gram-Negative Bacteria (1/3)

β -Lactamase	Examples	Substrates	Inhibition by Clavulanic Acid*	Molecular Class
Broad-spectrum	TEM-1, TEM-2, SHV-1	Benzylpenicillin (penicillin G), aminopenicillins (amoxicillin and ampicillin), carboxypenicillins (carbenicillin and ticarcillin), ureidopenicillin (piperacillin), narrow-spectrum cephalosporins (cefazolin, cephalothin, cefamandole, cefuroxime, and others)	+++	A
	OXA family	Substrates of the broad-spectrum group plus cloxacillin, methicillin, and oxacillin	+	D





Selected β -Lactamases of Gram-Negative Bacteria (2/3)

β -Lactamase	Examples	Substrates	Inhibition by Clavulanic Acid*	Molecular Class
Expanded-spectrum	TEM family and SHV family	Substrates of the broad-spectrum group plus oxyimino-cephalosporins (cefotaxime, cefpodoxime, ceftazidime, and ceftriaxone) and monobactam (aztreonam)	++++	A
	Others (BES-1, GES/IBC family, PER-1, PER-2, SFO-1, TLA-1, VEB-1, and VEB-2)	Same as for TEM family and SHV family	++++	A
	CTX-M family	Substrates of the expanded-spectrum group plus, for some enzymes, cefepime	++++	A





Selected β -Lactamases of Gram-Negative Bacteria (3/3)

β -Lactamase	Examples	Substrates	Inhibition by Clavulanic Acid*	Molecular Class
AmpC	ACC-1, ACT-1, CFE-1, CMY family, DHA-1, DHA-2, FOX family, LAT family, MIR-1, MOX-1, and MOX-2	Substrates of expanded-spectrum group plus cephamycins (cefotetan, cefoxitin, and others)	0	C
Carbapenemase	IMP family, VIM family, GIM-1, and SPM-1	Substrates of the expanded-spectrum group plus cephamycins and carbapenems (ertapenem, imipenem, and meropenem)	0	B
	KPC-1, KPC-2, and KPC-3	Same as for IMP family, VIM family, GIM-1, and SPM-1	+++	A
	OXA-23, OXA-24, OXA-25, OXA-26, OXA-27, OXA-40, and OXA-48	Same as for IMP family, VIM family, GIM-1, and SPM-1	+	D



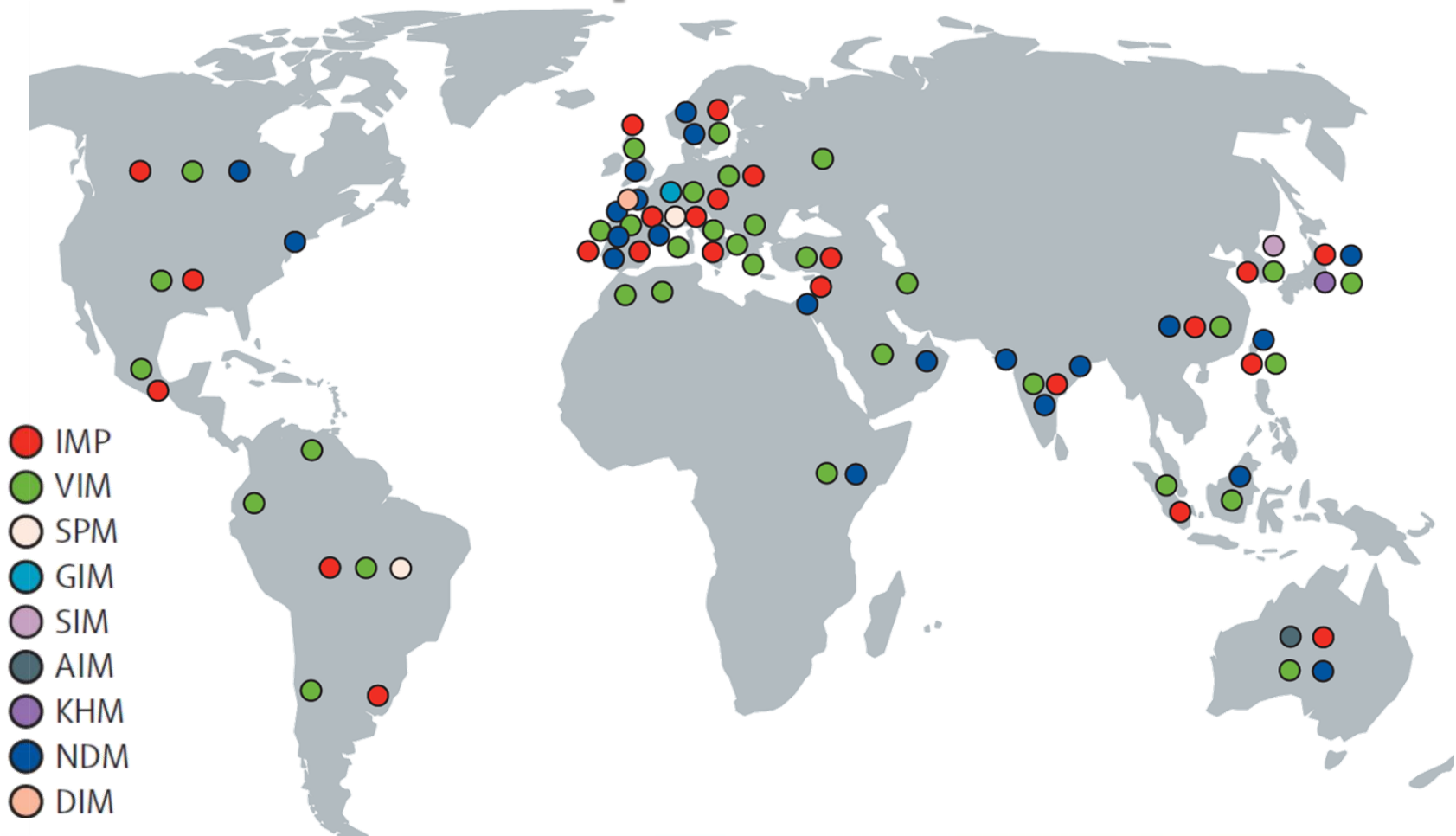
Metallo- β -Lactamases

IMP-1		VIM-2	
Japan	<i>Pseudomonas aeruginosa</i> ⁷	France	<i>P aeruginosa</i> ³⁵
Japan	Enterobacteriaceae ⁸	Korea	<i>A baumannii</i> ³⁶
Japan	<i>Acinetobacter baumannii</i> ⁹	Taiwan	Enterobacteriaceae ³⁷
Japan	<i>Acinetobacter xylosoxidans</i> ¹⁰	Greece	<i>A xylosoxidans</i> ³⁸
IMP-2		VIM-3	
Italy	<i>A baumannii</i> ¹¹	Taiwan	<i>P aeruginosa</i> ³⁹
Japan	<i>P aeruginosa</i> ¹⁰	Taiwan	<i>A baumannii</i> ³⁰
IMP-8		VIM-11	
Taiwan	Enterobacteriaceae ¹⁸	Argentina	<i>P aeruginosa</i> ⁵⁰
China	<i>A baumannii</i> ¹⁹	Taiwan	<i>A baumannii</i> ⁵¹
Portugal	<i>Pseudomonas mendocina</i> ²⁰	Taiwan	Enterobacteriaceae ⁵¹

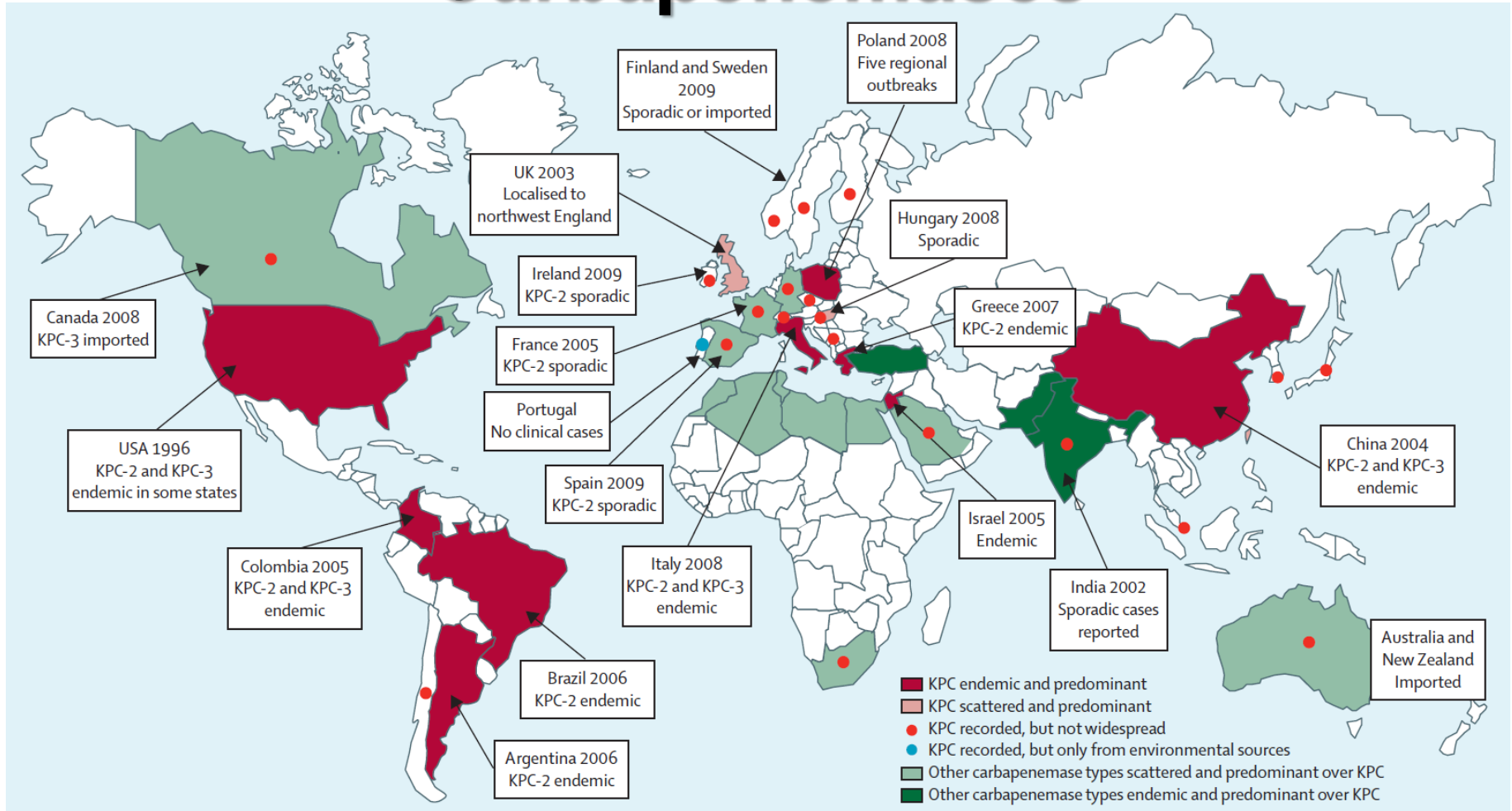
Carbapenem-resistant Enterobacteriaceae

- 同時為ESBL-producing (CTX-M)，並帶有outer membrane change。
- 同時為ESBL-producing (CTX-M)，並帶有up-regulation of efflux pump。
- 同時為AmpC β -lactamase-producing，並帶有outer membrane change。
- 同時為AmpC β -lactamase-producing，並帶有up-regulation of efflux pump。
- Secretion of carbapenemases in the plasmid.

Worldwide Types of Metallo- β -Lactamases



Global Expansion of KPC Carbapenemases





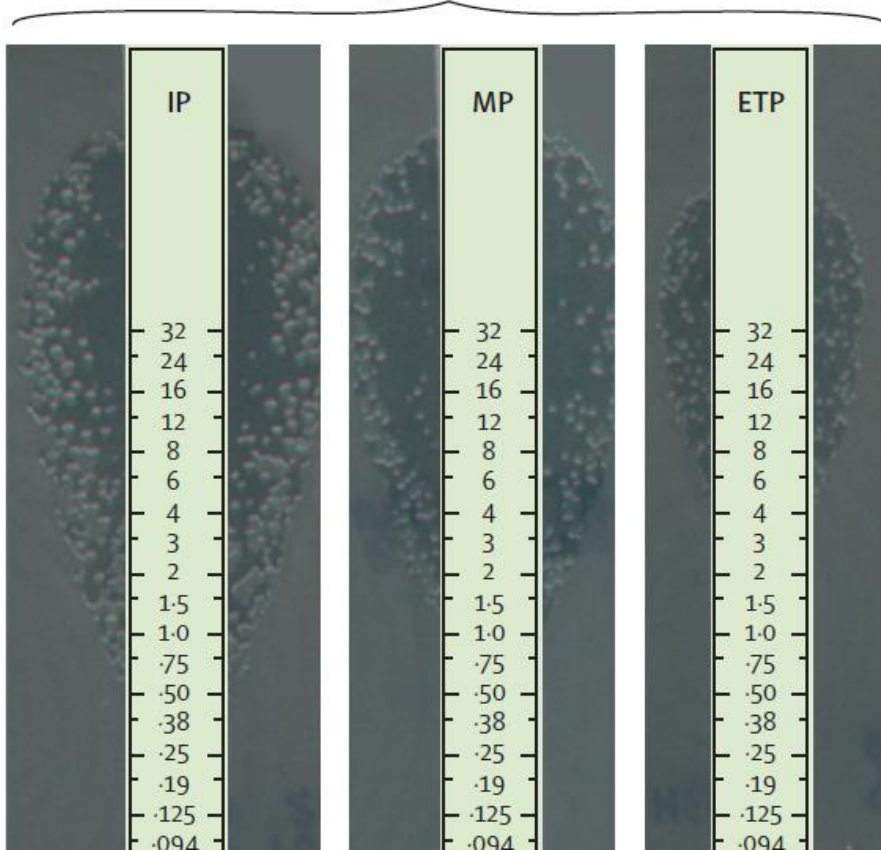
Mechanisms Involving Carbapenem Resistance of AB

- Metallo- β -lactamases (MBLs)
 - IMP-, VIM- group
- Carbapenem-hydrolysing oxacillinases (CHDLs)
 - OXA- group
- Loss of outer membrane proteins (OMPs [porin])
 - Decreased permeability
- Over expression of efflux pump

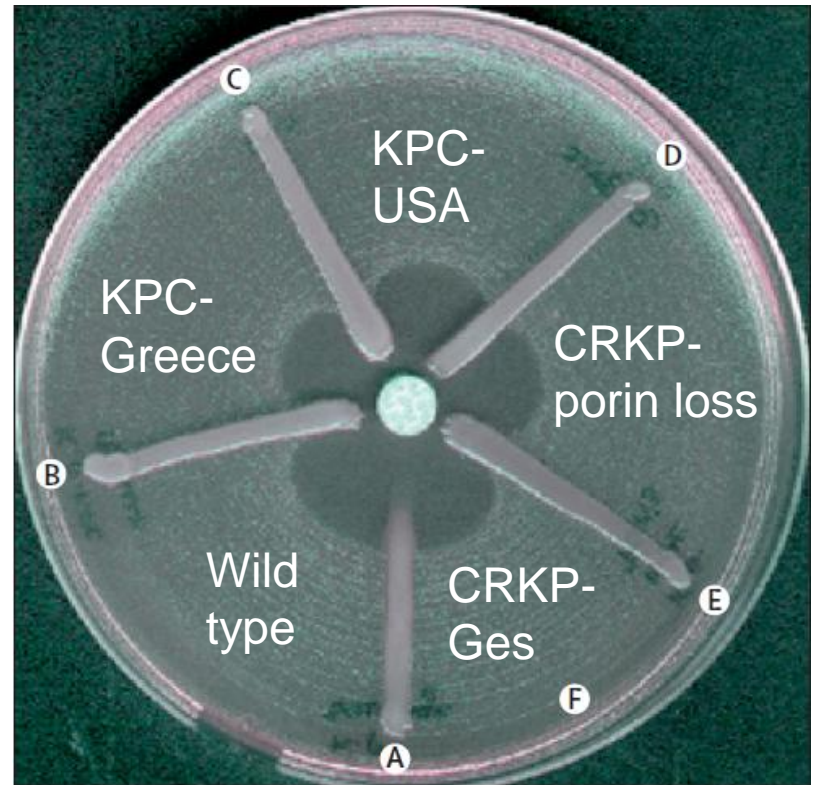


KPC-Enterobacteriaceae

K pneumoniae



Susceptibility- Etest



Modified Hodge test



Detection Carbapenemase in CRAB

Name	Nucleotide sequence (5'-3')	Location
Ab-GF	ACAAGAAATCTGCTCGT	<i>gyrA</i>
Ab-GR	CGAAGTTACCCTGACCATC	<i>gyrA</i>
5'-CS	GGCATCCAAGCAGCAAG	Integron
3'-CS	AAGCAGACTTGACCTGA	Integron
ACI10	GCTGAACGCGATAAACTTC	<i>ISAbal</i> of <i>bla</i> _{AmpC}
ACI2	TAGTACTGCTATTTACGGCT	<i>bla</i> _{AmpC}
IMP-A	GAAGGYGTTTATGTTCATAC	<i>bla</i> _{IMP}
IMP-B	GTAMGTTTCAAGAGTGATGC	<i>bla</i> _{IMP}
VIM2004A	GTTTGGTCGCATATCGCAAC	<i>bla</i> _{VIM}
VIM2004B	AATGCGCAGCACCAGGATAG	<i>bla</i> _{VIM}
OXA23-F	GATCGGATTGGAGAACCAGA	<i>bla</i> _{OXA-23}
OXA23-R	ATTTCTGACCGCATTTCCAT	<i>bla</i> _{OXA-23}
OXA24-F	GGTTAGTTGGCCCCCTTAAA	<i>bla</i> _{OXA-24-like}
OXA24-R	AGTTGAGCGAAAAGGGGATT	<i>bla</i> _{OXA-24-like}
OXA51-F	TAATGCTTTGATCGGCCTTG	<i>bla</i> _{OXA-51-like}
OXA51-R	TGGATTGCACTTCATCTTGG	<i>bla</i> _{OXA-51-like}
OXA-58-F	AAGTATTGGGGCTTGTGCTG	<i>bla</i> _{OXA-58}
OXA-58-R	CCCCTCTGCGCTCTACATAC	<i>bla</i> _{OXA-58}
OXA-58A	CGATCAGAATGTTCAAGCGC	<i>bla</i> _{OXA-58}
OXA-58B	ACGATTCTCCCCTCTGCGC	<i>bla</i> _{OXA-58}
ISAb3B	CGTTTACCCCAAACATAAGC	<i>tnpA</i> of <i>ISAb3</i> (but not in <i>ISAb3-like</i>)
ISAb3C	AGCAATATCTCGTATACCGC	<i>tnpA</i> of <i>ISAb3-like</i> and <i>ISAb3</i>
ISAb1-F	GGATCCCTCTGTACACGAYAAATTC	<i>ISAb1</i>
OXA23-R1	GAATTCTTAAATAATATTCAGCTGTTTAAATG	<i>bla</i> _{OXA-23}
OXA51-R1	GAATTCCTATAAAAATACCTAATTGTTCTAAAC	<i>bla</i> _{OXA-51-like}

資料來源：Lin YC, Sheng WH, et al. Antimicrob Agents Chemother 2010; 54:2078-84

Main Mechanisms of Antimicrobial Resistance of *A. baumannii*

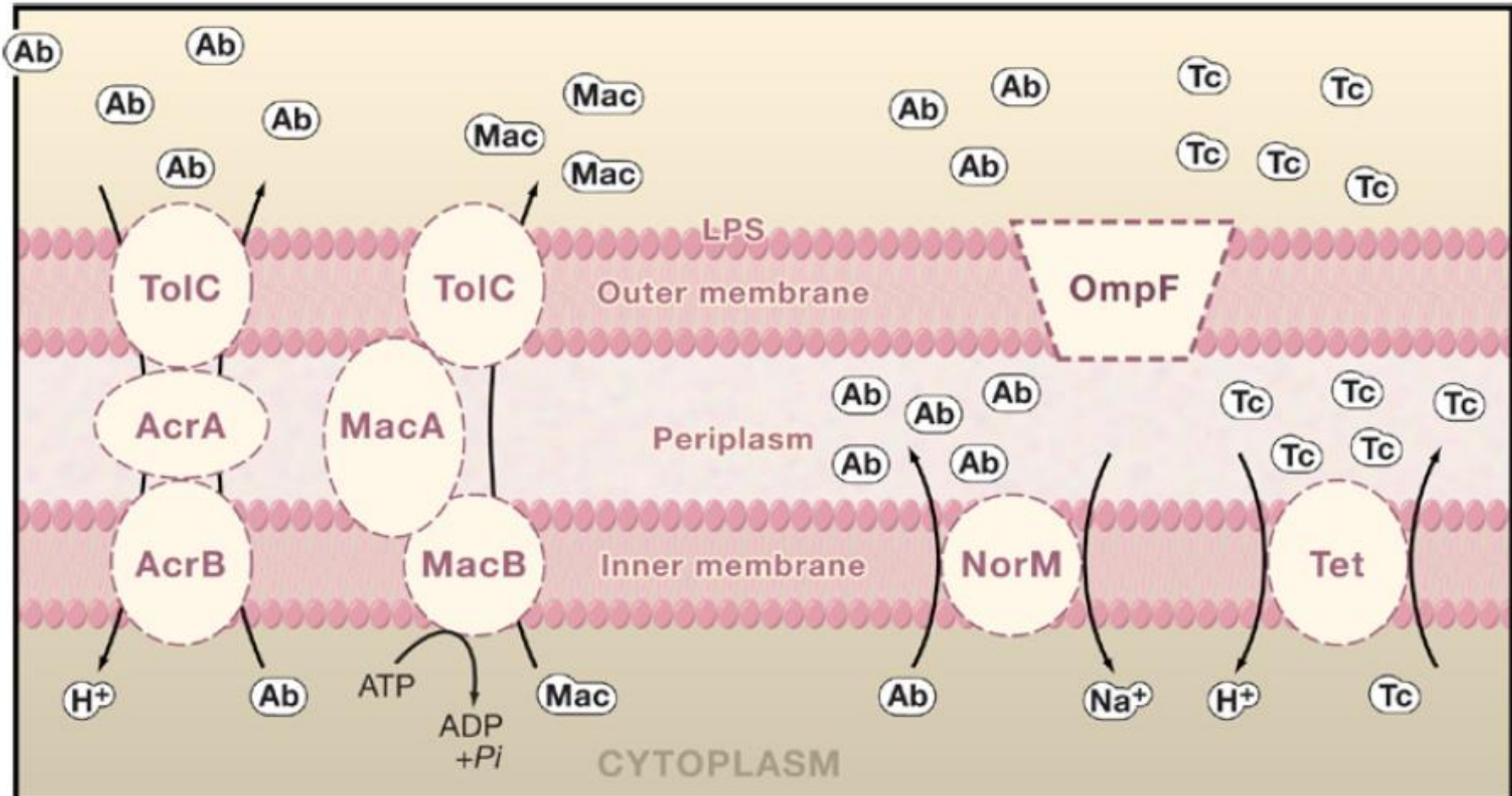
Remarkable capacity to show resistance to antibiotics of most classes

- Intrinsically, or
- Following acquisition of resistance

Various mechanisms of resistance

- Production of β lactamases
- Efflux pumps
- Lower permeability of the outer membrane
- Mutations in antibiotic targets (eg, for quinolones)
- Production of enzymes inactivating aminoglycosides

Efflux Pump and Outer Membrane Protein





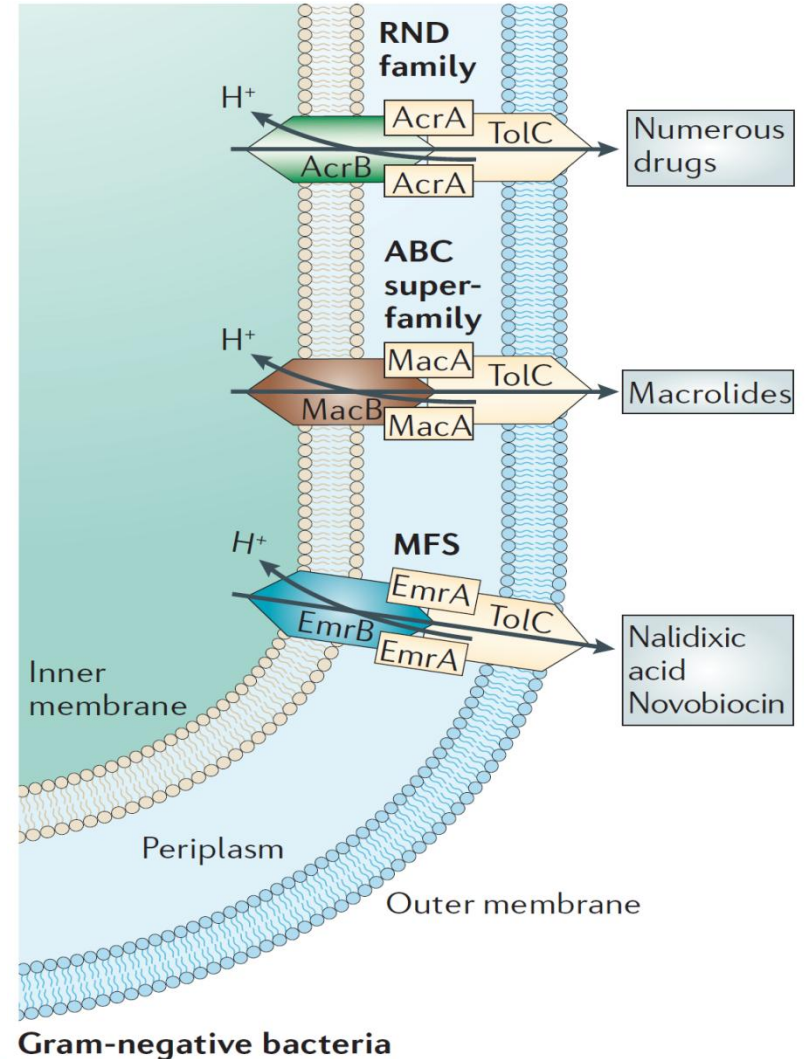
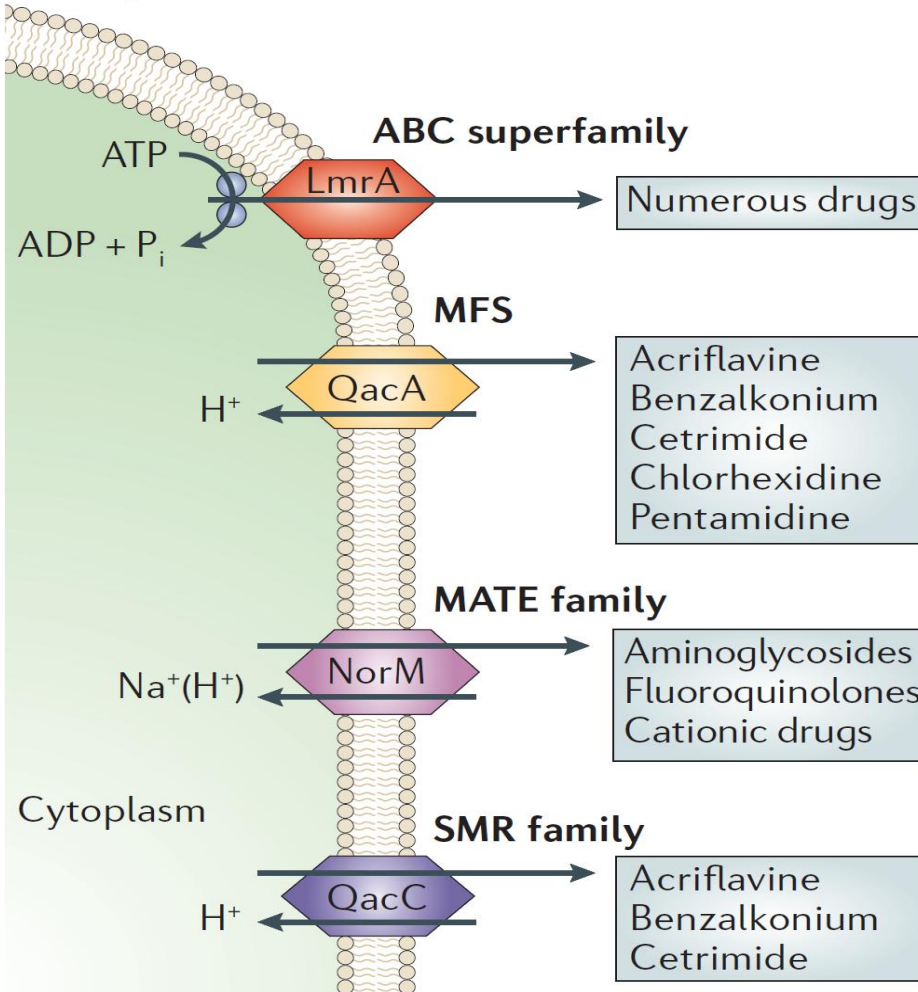
Families of Multidrug-resistant Efflux Pumps

- ATP-binding cassette (ABC) superfamily
- Major facilitator superfamily (MFS)
- Multidrug and toxic-compound extrusion (MATE) family
- Small multidrug resistance (SMR) family
- Resistance nodulation division (RND) family



Multidrug-resistant Efflux Pumps

Gram-positive bacteria





5. 結論

結論

- 抗藥性細菌增加與抗生素大量使用有關。
- 細菌發展不同抗藥機轉的演進，可導致細菌對不同類抗生素均有抗藥性。
- 抗藥性基因可以利用多種機制（如質體或轉位子）在不同細菌間傳遞。
- 細菌可以同時存在抗藥性基因於染色體或質體上於抗生素環境時使用。



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